

Stomatal and mesophyll conductances control CO₂ transfer to chloroplasts in leaves of grapevine (*Vitis vinifera* L.)

H. DÜRING

Institut für Rebenzüchtung Geilweilerhof der Bundesanstalt für Züchtungsforschung an Kulturpflanzen, Siebeldingen, Deutschland

Summary

From simultaneous determination of net CO₂ assimilation and transpiration at the abaxial side and of the photosynthetic electron transport rate at the adaxial side of field-grown, light-saturated leaves of grapevine (cv. Riesling) photorespiration, stomatal conductance for CO₂, mesophyll conductance and the CO₂ concentration in intercellular spaces (C_i) and in chloroplasts (C_c) were estimated. CO₂ assimilation was saturated at about C_i = 340 ppm. At increasing ambient CO₂ concentration (C_a) photorespiration decreased (less negative values); stomatal conductance decreased significantly (-45 %) limiting CO₂ uptake into intercellular spaces. Rates of total photosynthetic electron transport were constant between C_i = 340 and 800 ppm and decreased by 34 % at low C_i. Electron flow to carboxylation was closely correlated to CO₂ assimilation rates (R² = 0.999). When C_a was raised, the CO₂ concentration in chloroplasts (C_c) increased but at smaller rates than C_i. Presumably due to the distinct decline of the mesophyll conductance C_c remained constant at C_i > 340 ppm. At C_a = 400 ppm the C_c/C_a ratio was 0.46–0.48, corroborating data reported for other species (CORNIC and FRESNEAU 2002). At 2 % ambient O₂ and 400 ppm CO₂ decreased rates of photorespiration (-69 %) were associated with a decline of total photosynthetic electron flow (-6 %); higher stomatal and mesophyll conductances, however, led to increases of C_c and CO₂ assimilation rates (+49 %). It is hypothesized that both stomatal and mesophyll conductance are involved in the adaptation of the CO₂ supply to the CO₂ demand at the site of carboxylation in chloroplasts.

Key words: photosynthesis, photorespiration, photosynthetic electron transport, chloroplastic carbon dioxide, stomatal conductance, mesophyll conductance.

Introduction

Leaves of grapevines are characterised by relative low rates of CO₂ assimilation (generally $\leq 20 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$) under saturating light, ambient CO₂ concentration and favourable air humidity and temperature condition as compared to some herbaceous plants with values up to $40 \mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ (LARCHER 1975, DÜRING 1991, EPRON *et al.* 1995).

This characteristic of woody plants may be associated with biochemical constraints on CO₂ assimilation, *i.e.* Rubisco activity or chloroplast capacity for electron trans-

port and/or limitations of the CO₂ transfer rate from the ambient air to the chloroplasts (LLOYD *et al.* 1992).

Stomatal conductance for CO₂ (g_{CO₂}) of grape leaves generally varies between 0 and 200 mmol m⁻² s⁻¹ and, at ambient CO₂ concentration (350–380 ppm), intercellular CO₂ concentration (C_i) ranges from 250 to 300 ppm. These values are not different from other C₃ plants (LLOYD *et al.* 1992). According to BOTA *et al.* (2002) the ribulose 1,5-bisphosphate carboxylase/oxygenase (Rubisco) activity in leaves of field-grown grapevine (cv. Tempranillo) was close to the light- and CO₂-saturation rate of photosynthesis and in our experiments with field-grown grape varieties total electron transport rates (J_p) ranged from 80 to 190 $\mu\text{mol e}^- \text{ m}^{-2} \text{ s}^{-1}$ (ORTOIZZE and DÜRING 2001). Since these values are well above those required for CO₂ assimilation (LLOYD *et al.* 1992), it has been suggested that CO₂ assimilation in woody plants may be limited by low internal ('mesophyll') conductance for CO₂ (g_{mes}), *i.e.* the diffusion of CO₂ through mesophyll cell walls, the plasmalemma, part of the cytosol, and the chloroplast envelope and some of the chloroplast stroma (NOBEL 1983, EPRON *et al.* 1995, LAWLOR 2002).

LAWLOR and CORNIC (2002) assumed that under optimum ambient conditions in fully hydrated leaves CO₂ assimilation can be increased to the potential rate of photosynthesis (A_{pot}) by increasing C_a to 800 ppm, on the premise that "g_m (equals g_{mes}, the author) is not changed when C_a increases"; thus, at increasing C_a and C_i changes of the CO₂ transfer in the mesophyll can not be excluded *a priori*.

The aim of the present study was to elucidate the CO₂ transfer in leaves of irrigated grapevines by determining stomatal and mesophyll conductances as well as intercellular and chloroplastic CO₂ concentrations as a function of increasing ambient CO₂ concentration.

Material and Methods

In June 2002 the distal end of shoots from field-grown Riesling vines with 6–8 fully expanded leaves were cut and immediately re-cut under water to avoid air embolism in xylem vessels. During and after transfer to the laboratory the shoots were kept in water. Light was provided by two lamps (Powerstar HQI-T, 400 W/DH, Osram). The leaf water potential under these conditions ranged between -0.25 and -0.35 Mpa.

Combined gas exchange and chlorophyll fluorescence measurements: Fully expanded leaves were used to measure simultaneously gas

exchange and chlorophyll fluorescence at the same leaf segment using a 'HCM-1000-Photosynthesis-System' (Walz, Effeltrich, Germany) combined with a 'Mini-PAM-System' (Walz, Germany).

Leaves were inserted into the cuvette to measure gas exchange at their abaxial side and simultaneously chlorophyll fluorescence (quantum yield) at the adaxial side.

Measurements were performed at light-saturation of CO₂ assimilation (750 μmol m⁻² s⁻¹) between 8 and 11 a.m. Leaf temperature was kept constant at 25 °C, relative air humidity in the cuvette at 50-55 %. CO₂ concentration of the gas stream leading to the measuring cuvette with the inserted leaf was increased stepwise from 50 to 1000 or 2000 ppm CO₂, respectively. The net CO₂ assimilation rate (A), the stomatal conductance for CO₂ (g_{CO2}) and the intercellular CO₂ concentration (Ci) were calculated according to VON CAEMMERER and FARQUHAR (1981) and the quantum yield of the illuminated leaf area (Fv/Fm') was determined by saturating light pulses as described earlier (DÜRING 1998, DÜRING and DAVTYAN 2001).

According to KRALL and EDWARDS (1992) the total electron flow (J_t) can be derived from the quantum yield of PSII (Y), the light intensity incident on the leaf (PAR), the fractional absorptance of light by the leaf (a) and the absorptance by PSII divided by the absorptance of PSI + PSII (f):

$$J_t = Y \cdot PAR \cdot a \cdot f,$$

where 'a' equals 0.84 and 'f' equals 0.5 (SCHREIBER 1997).

J_t can be divided into its components, J_c and J_o, the electron flow rates devoted to carboxylation and oxygenation of RuBP according to EPRON *et al.* (1995):

$$J_c = 1/3 [J_t + 8(A + R_d)] \text{ and } J_o = 2/3 [J_t - 4(A + R_d)],$$

where A is the CO₂ assimilation rate and R_d the dark respiration. R_d was determined by gas exchange measurements after keeping leaves in the dark for 10 min.

Photorespiration (R_L) was calculated according to VALENTINI *et al.* (1995):

$$R_L = 1/12 [J_t - 4(A + R_d)]$$

The CO₂ concentration at the chloroplast level, C_c, was determined according to CORNIC and FRESNEAU (2002) assuming a specificity factor of 80, which is commonly used for C₃ plants:

$$C_c = 2\Gamma (J_c / J_o),$$

where $\Gamma = 42.5$ ppm at a leaf temperature of 25 °C. Meanwhile BOTA *et al.* (2002) have reported somewhat lower specificity factors for leaves of grapevine, the value depending, *inter alia*, on variety and leaf age.

The mesophyll (or 'internal') conductance (g_{mes}) was calculated as proposed by HARLEY *et al.* (1992):

$$g_{mes} = A / (C_i - C_c)$$

Results presented are mean values of 4-5 replicates, bars denote confidence limits at the 5 % level.

Results and Discussion

Under saturating light conditions CO₂ assimilation rates (A) of Riesling leaves increased with increasing intercellular CO₂ concentration (Ci) induced by stepwise increases of ambient CO₂ concentration (Ca) (Fig. 1). Photosynthesis was almost CO₂-saturated at Ci = 340 and increased only slightly

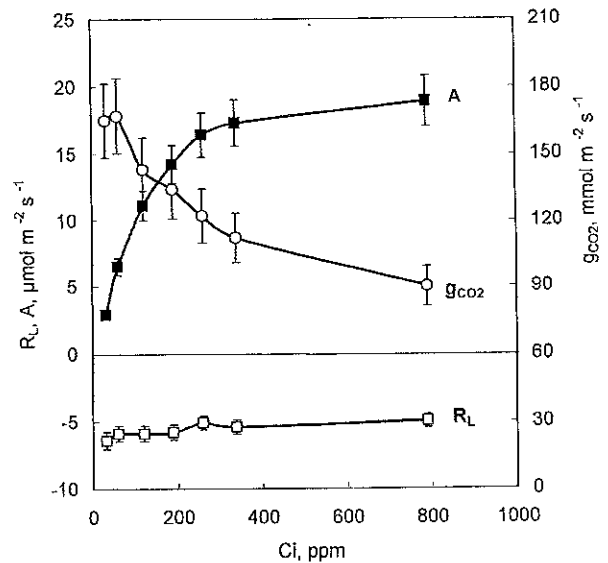


Fig. 1 CO₂ assimilation (A), stomatal conductance for CO₂ (g_{CO2}) and photorespiration (R_L) as a function of intercellular CO₂ concentration (Ci). Bars denote confidence limits at the 5 % level.

at higher Ci. Concurrently photorespiration (R_L) decreased by 29 % (values less negative) and stomatal conductance (g_{CO2}) by 45 %.

It has been shown earlier that stomatal pore size of grapevine leaves declines at increasing Ca, the sensitivity of stomata to CO₂ being higher in dehydrated leaves (DÜRING 1991). According to RASCHKE (1975) the stomatal response to CO₂ can be ascribed to increases of CO₂ in the substomatal cavity and the pore, *i.e.* to Ci rather than to Ca.

The total electron transport rate (J_t) and its component devoted to carboxylation, J_c, were constant at high Ci (340 and 800 ppm) and decreased by 34 % at low Ci (Fig. 2); J_c was closely related to rates of CO₂ assimilation (Fig. 3). Interestingly, at low Ci (32 ppm) A was close to zero while J_t decreased only by 34 %.

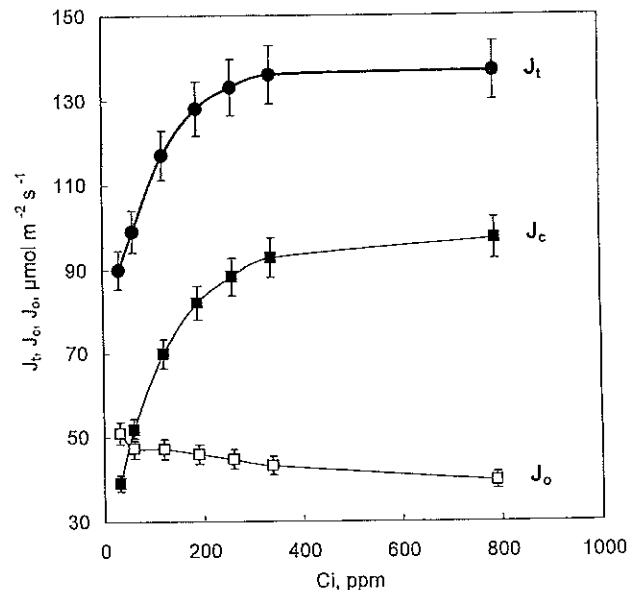


Fig. 2 Total electron flow (J_t), electron flow to carboxylation (J_c) and to oxygenation (J_o) as a function of the intercellular CO₂ concentration (Ci). For details see Fig. 1.

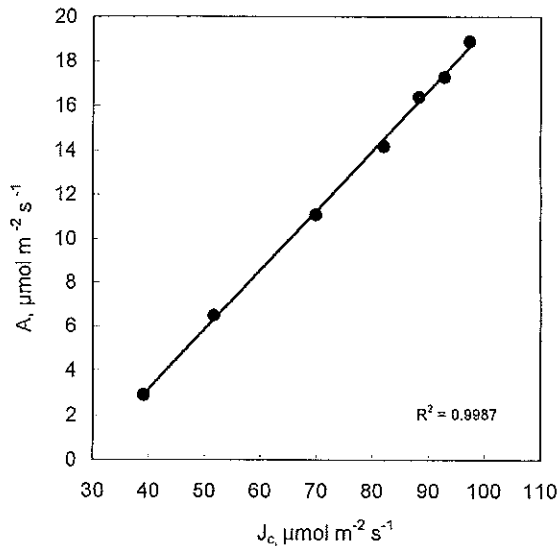


Fig. 3 The relationship between CO_2 assimilation (A) and the electron flow to carboxylation (J_c).

Similar results have been reported by CORNIC and BRIANTAIS (1991) and CORNIC and FRESNEAU (2002) who concluded that the photosynthetic carbon reduction and carbon oxidation cycles are the main electron sinks for photosystem II activity during a mild drought, *i.e.* when CO_2 supply to chloroplasts is limited. Obviously, under these conditions oxygen can efficiently replace carbon dioxide as an electron acceptor. In their experiments at low C_i or in dehydrated leaves the allocation of photosynthetic electrons to O_2 was increased. This corroborates our data indicating that at $C_i = 32$ ppm electron flow to O_2 reduction (J_o) was higher than the electron flow to CO_2 reduction (J_c) (Fig. 2).

The relationship between A and C_c is shown in Fig. 4. The initial linear slope of the curve represents the carboxylation efficiency, *i.e.* the stage where RuBP is saturated (FARQUHAR and SHARKEY 1982). In the past the carboxylation efficiency was approximately derived from A/C_i relationships, however, under various stress conditions estimation of C_i in leaves of grapevine (and some other 'heterobaric' species) is misleading due to the heterogenous behaviour of stomata ('stomatal patchiness') (review: WEYERS and LAWSON 1997, grapevine: DÜRING and LOVEYS 1996). Relating A to C_c instead of C_i appears to be a most promising way to by-pass the problem and to determine carboxylation efficiency at the site of carboxylation, *i.e.* more accurately.

The almost linear increase of C_i with C_a (Fig. 5) demonstrates that CO_2 uptake into substomatal cavities and intercellular spaces was limited only to a small extent by stomata, *e.g.* at $C_a = 1000$ ppm C_i was reduced only by 21 %.

Like C_i , C_c increased when C_a was raised, however, at somewhat smaller rates; *e.g.*, at $C_a = 400$ ppm C_c was lower than C_i (-77 ppm or -30 %). This is in line with observations of KRALL and EDWARDS (1992) who assumed a 10 % decrease in CO_2 in the chloroplasts relative to the concentration in the substomatal cavity "due to diffusive resistance in the aqueous phase" and considerably higher values for leaves of woody plants. Similar results were obtained from 13 woody plants by EPRON *et al.* (1995) who concluded that "the low values of net CO_2 uptake that occur in some woody plants

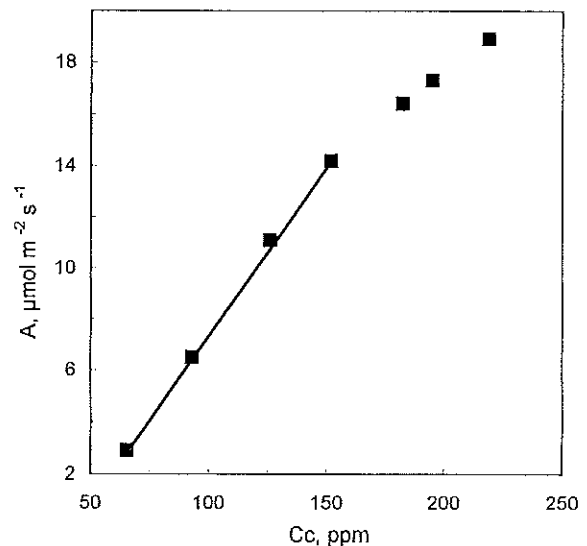


Fig. 4 The relationship between CO_2 assimilation (A) and the chloroplast CO_2 concentration (C_c).

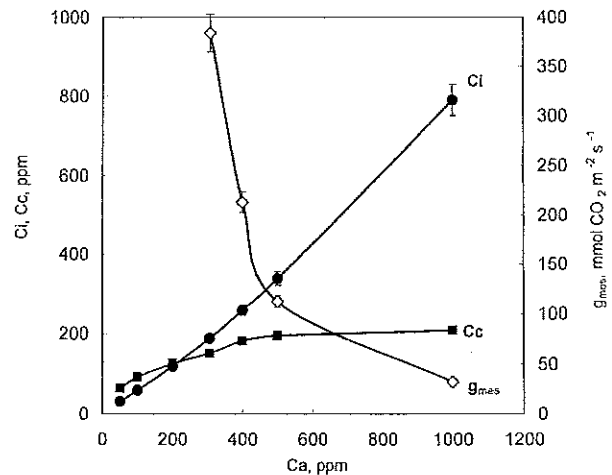


Fig. 5 The intercellular CO_2 concentration (C_i), the chloroplast CO_2 concentration (C_c) and the mesophyll conductance (g_{mes}) as a function of ambient CO_2 concentration (C_a). For details see Fig. 1.

are partly ascribable to high internal resistances". To the best of my knowledge, the only data for grape, obtained however by a different methodological approach, are those of FLEXAS *et al.* (2002) indicating that in irrigated vines C_c was only about 50-60 % of C_i .

In our trials, at $C_a = 400$ ppm the ratio C_c/C_a was 0.46-0.48 which is fairly close to the values reported by CORNIC and FRESNEAU (2002).

At $C_a > 500$ ppm, C_c no longer increased, leading to an increasing divergence between values of C_i and C_c . Concurrently, g_{mes} declined significantly (-92 %).

When, in similar experiments, C_a values were raised to 2.000 ppm, C_i increased to 1.600 ppm while, again, C_c values increased at smaller rates reaching only about 10 % of the final C_i values. In parallel, g_{mes} declined about 6-fold reflecting a severe increase of the mesophyll resistance to the CO_2 transfer at high CO_2 supply. Meanwhile, responses of C_c and g_{mes} to alterations of ambient C_a and C_i have been confirmed for other *Vitis vinifera* cvs and *Helianthus annuus* (DÜRING, unpubl.).

These results indicate that increases of ambient CO₂ supply affect stomatal and mesophyll conductance to different degrees, thereby controlling the CO₂ transfer to the intercellular spaces and chloroplasts. It is hypothesised that both, stomatal and mesophyll conductance adapt the CO₂ supply to the actual CO₂ demand in chloroplasts.

This view is supported by experiments in which the CO₂ demand was raised by lowering ambient O₂ concentration from 21 % to 2 % (Table). As expected, photorespiration declined significantly (-69 %) leading to a small reduction of total photosynthetic electron flow. Similar results were obtained by CORNIC and BRIANTAIS (1991) reporting a decrease of electron transport at 1 % ambient O₂. Our data indicate that under non-photorespiratory conditions the increased demand for CO₂ was associated with higher stomatal and mesophyll conductances leading to increased CO₂ concentrations in chloroplasts.

Table

Photosynthetic parameters under photorespiratory (21 % O₂) and non-photorespiratory (2 % O₂) conditions. Ambient CO₂ concentration: 400 ppm, leaf temperature: 25 °C, photosynthetic active radiation: 750 μmol m⁻² s⁻¹ (cv. Riesling)

Oxygen, %	g_{CO_2}	Ci	g_{mes}	Cc	A	R_L	J_t
21	102	291	104	190	10.5	3.10	81
2	167	299	205	223	15.6	0.97	76

g_{CO_2} , g_{mes} (mmol m⁻² s⁻¹); Ci, Cc (ppm); A, R_L , J_t (μmol m⁻² s⁻¹). See Material and Methods.

Conclusion

The presented data indicate that the relative low maximal CO₂ assimilation rates of grapevine leaves may, in fact, be explained by their relative high Ci-Cc difference, *i.e.* their high mesophyll resistance for CO₂. Moreover, the premise of LAWLOR and CORNIC (2002) - see Introduction - has been verified, since, in fact, increasing Ca altered mesophyll conductance. The data also raise questions, *e.g.* as to the site and mechanism of CO₂ sensing in leaves and the control of CO₂ transfer in the mesophyll. Determination of chloroplastic CO₂ concentration in grapevine genotypes will enable estimation of variety specific carboxylation efficiency and, for dehydrated vines, may contribute to elucidate causal relationships of factors limiting CO₂ assimilation.

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