



# Whole-Genome Sequencing of the Fungus *Penicillium citrinum* Reveals the Biosynthesis Gene Cluster for the Mycotoxin Citrinin

Markus Schmidt-Heydt,<sup>a</sup> Dominic Stoll,<sup>a</sup> Rolf Geisen<sup>a</sup>

<sup>a</sup>Department of Safety and Quality of Fruit and Vegetables, Max Rubner-Institut, Federal Research Institute of Nutrition and Food, Karlsruhe, Germany

**ABSTRACT** *Penicillium citrinum* is a food-contaminating ascomycete that consistently produces large amounts of the mycotoxin citrinin. Citrinin exhibits, besides its toxicity, antibiotic effects and thus potentially forces antibiotic resistance. Within the genome sequence, we identified the biosynthesis gene cluster for citrinin, which appears to be highly conserved within the genus *Penicillium*.

*Penicillium citrinum* occurs on salt-rich products, citrus fruits, and wheat and other cereals, the presence of which leads to contamination with the mycotoxin citrinin, and *P. citrinum* is related to the species *P. expansum*, *P. nordicum*, and *P. verrucosum* (1–3). Citrinin has detrimental effects on the kidneys and the immune system (4). Moreover, the first statin, mevastatin, was isolated from *P. citrinum* in the 1970s (5).

Genome sequencing was carried out on the MiSeq platform (Illumina) as follows: genomic DNA of *P. citrinum* strain DSM 1997 was extracted from a pure culture, after growing for 7 days at 25°C on malt extract sucrose agar slants, with the NucleoSpin Plant II kit (Macherey-Nagel) and then quantified and quality checked with NanoDrop 1000 (VWR International) and Qubit 3.0 spectrometers/fluorometers. The sequencing library was built with the Illumina Nextera XT DNA kit and quality checked with the Experion DNA 1K analysis kit (Bio-Rad Laboratories). Raw reads (read length, 2 × 300 bp) were processed with the FastQ preprocessing toolkit (Blast2GO Pro v.5.2). *De novo* assembly was done with SeqMan NGen v.12.3 (Lasergene) with default settings; contigs smaller than 200 nucleotides (nt) and mitochondrial sequences were removed. The assembly size was 31,529,786 bp with 64× coverage and 976 genomic contigs; other parameters were an  $N_{50}$  value of 67,438 kb, an  $L_{50}$  value of 137 kb, and a G+C content of 46.15% ± 2.84%. Prediction of biosynthesis gene clusters (BGCs) was carried out with antiSMASH fungal v.3.0 with the cluster-finder algorithm for BGC border prediction with default settings (6, 7).

Twenty-nine BGCs were predicted, 9 type 1 polyketide synthase (T1PKS) clusters, 3 nonribosomal peptide synthetase (NRPS) clusters, 2 polyketide synthase (PKS)-NRPS hybrid clusters, 1 indole-NRPS hybrid cluster, 2 fatty-acid clusters, 1 terpene cluster, and 11 BGCs which were not specified further. Genome annotation based on *Aspergillus nidulans* was done with AUGUSTUS v.3.0.2 (8), and we identified 9,754 genes.

Within the genome sequence of *P. citrinum* DSM 1997 a complete BGC for citrinin, showing homology and a high sequence similarity to the citrinin cluster of *Monascus purpureus* (9), *P. expansum*, and *P. verrucosum* (10, 11), was identified. Future analyses of the genome sequence of this food-relevant fungal species will give deeper insight into the secondary metabolite biosynthesis of *P. citrinum* compared to that of other citrinin-producing fungi.

**Data availability.** The whole-genome sequence of *P. citrinum* strain DSM 1997 has been deposited at NCBI under the accession number [LKUP00000000](https://ncbi.nlm.nih.gov/submit/submitseq/submitseq.cgi?accession=LKUP00000000), and the raw reads were deposited in the SRA under the accession number [PRJNA298119](https://ncbi.nlm.nih.gov/submit/submitseq/submitseq.cgi?accession=PRJNA298119).

**Citation** Schmidt-Heydt M, Stoll D, Geisen R. 2019. Whole-genome sequencing of the fungus *Penicillium citrinum* reveals the biosynthesis gene cluster for the mycotoxin citrinin. *Microbiol Resour Announc* 8:e01419-18. <https://doi.org/10.1128/MRA.01419-18>.

**Editor** Antonis Rokas, Vanderbilt University

**Copyright** © 2019 Schmidt-Heydt et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Markus Schmidt-Heydt, [Markus.Schmidt-Heydt@mri.bund.de](mailto:Markus.Schmidt-Heydt@mri.bund.de).

**Received** 19 October 2018

**Accepted** 12 December 2018

**Published** 24 January 2019

## ACKNOWLEDGMENTS

This work was funded by the Brigitte and Wolfram Gedek Foundation and the Max Rubner-Institut.

We thank Anastasia Landeis and Karla Hell for excellent technical assistance.

## REFERENCES

1. Houbraken JAMP, Frisvad JC, Samson RA. 2010. Taxonomy of *Penicillium citrinum* and related species. *Fungal Divers* 44:117–133. <https://doi.org/10.1007/s13225-010-0047-z>.
2. Pitt JI, Hocking AD. 2009. *Fungi and food spoilage*. Springer Nature, Heidelberg, Germany.
3. Malmstrøm J, Christophersen C, Frisvad JC. 2000. Secondary metabolites characteristic of *Penicillium citrinum*, *Penicillium steckii* and related species. *Phytochemistry* 54:301–309. [https://doi.org/10.1016/S0031-9422\(00\)00106-0](https://doi.org/10.1016/S0031-9422(00)00106-0).
4. Föllmann W, Behm C, Degen GH. 2014. Toxicity of the mycotoxin citrinin and its metabolite dihydrocitrinone and of mixtures of citrinin and ochratoxin A in vitro. *Arch Toxicol* 88:1097–1107. <https://doi.org/10.1007/s00204-014-1216-8>.
5. Endo A, Kuroda M, Tsujita Y. 1976. ML-236A, ML-236B, and ML-236C, new inhibitors of cholesterol synthesis produced by *Penicillium citrinum*. *J Antibiot* 29:1346–1348. <https://doi.org/10.7164/antibiotics.29.1346>.
6. Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res* 43:W237–W243. <https://doi.org/10.1093/nar/gkv437>.
7. Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences. *Nucleic Acids Res* 39:W339–W346. <https://doi.org/10.1093/nar/gkr466>.
8. Sommerfeld D, Lingner T, Stanke M, Morgenstern B, Richter H. 2009. AUGUSTUS at MediGRID: adaption of a bioinformatics application to grid computing for efficient genome analysis. *Future Gener Comput Syst* 25:337–345. <https://doi.org/10.1016/j.future.2008.05.010>.
9. Li Y-P, Xu Y, Huang Z-B. 2012. Isolation and characterization of the citrinin biosynthetic gene cluster from *Monascus aurantiacus*. *Biotechnol Lett* 34:131–136. <https://doi.org/10.1007/s10529-011-0745-y>.
10. Geisen R, Schmidt-Heydt M, Touhami N, Himmelsbach A. 2018. New aspects of ochratoxin A and citrinin biosynthesis in *Penicillium*. *Curr Opin Food Sci* <https://doi.org/10.1016/j.cofs.2018.04.001>.
11. Geisen R, Schmidt-Heydt M, Stoll D, Touhami N. 2018. Aspects of the occurrence, genetics, and regulation of biosynthesis of the three food relevant *Penicillium* mycotoxins: ochratoxin A, citrinin, and patulin, p 413–433. *In* Anke T, Schöffler A (ed), *The Mycota: a comprehensive treatise on fungi as experimental systems for basic and applied research. Physiology and genetics: selected basic and applied aspects*. Springer, Cham, Germany.