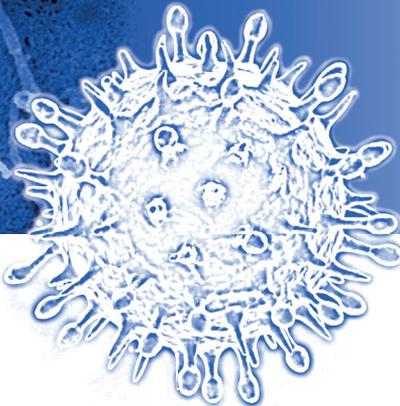


28th to 30th
September 2016

Alfried Krupp
Wissenschaftskolleg
Greifswald



1st SUMMER SCHOOL "INFECTION BIOLOGY"



Welcome note

Dear colleagues,

It is a great pleasure to welcome you to the 1st Summer School on "Infection Biology". This conference is a joint effort of the DFG Research Training Group 1870 "Bacterial Respiratory Infection - Common and Specific Mechanisms of Pathogen Adaptation and Immune Defense", the Friedrich-Loeffler-Institut (FLI), Federal Research Institute for Animal Health, and the Alfried Krupp Wissenschaftskolleg Greifswald.

This truly interdisciplinary event addresses established experts and promising young scientists working in microbiology, virology and infection biology. The topics include state-of-the-art presentations on bacterial and viral interactions with the human host as well as recent advances in the understanding of co-infection mechanisms and pathophysiology. The program of the Summer School combines keynote lectures, oral presentations by experts in the field and young investigators, workshops and posters by the PhD students of the RTG1870 and FLI. Moreover, a panel discussion on Ethics and the Dual-use issue in biological sciences will be held by prominent researchers. Additionally, several informal get-togethers will facilitate the scientific exchange between invited speakers and participants and aim to establish new contacts or collaborations.

We are sure that the historic environment of Greifswald - a former member of the Hanseatic League - combined with the modern aspects of a university town at the Baltic Sea will be the perfect place to create a stimulating atmosphere of exciting talks, inspiring discussions, and scientific curiosity. We are pleased to welcome you in Greifswald and wish you a successful and stimulating conference.

With kind regards,
Yours sincerely,



Prof. Dr. rer. nat.
Sven Hammerschmidt

Chair of the Department
Genetics of Microorganisms
and Speaker of the DFG-
GRK1870



Prof. Dr. rer. nat. Dr. med.
vet. h. c.
Thomas C. Mettenleiter

President of FLI, Insel Riems
Chair of the Institute of
Molecular Virology and Cell
Biology at FLI



Prof. Dr. rer. nat.
Bärbel Friedrich

Academic Director of the
Alfried Krupp Wissenschafts-
kolleg



Content

Program	7
Keynote Lecture	10
Session I: Respiratory Bacterial-Viral Co-Infection	12
Session II: Biosafety	20
Session III: Ethics and Dual use issues in the Biological Sciences	24
Session IV: Host-Pathogen-Interactions of Bacteria and Viruses	26
Session V: Imaging and OMICS technologies in the Infection Research	34
Poster	42

Program

Wednesday, September 28th, 2016

Venue: **Alfried Krupp Wissenschaftskolleg Greifswald**

- 18:00 **Welcome**
Bärbel Friedrich (Greifswald)
Sven Hammerschmidt (Greifswald)
Thomas C. Mettenleiter (Greifswald – Insel Riems)
- 18:15 **Keynote Lecture: Influenza: Virus-Host-Bacterium**
Hans-Dieter Klenk (Marburg)
- 19:00 Dinner

Thursday, September 29th, 2016

Session I: Respiratory Bacterial-Viral Co-Infection

Venue: **Alfried Krupp Wissenschaftskolleg Greifswald**

Chair: **Thomas C. Mettenleiter & Sven Hammerschmidt**

- 09:00 **Culture systems for differentiated airway epithelial cells and their infection by swine influenza viruses**
Georg Herrler (Hannover)
- 09:45 **Colonization and invasion strategies of Streptococcus suis in the porcine respiratory tract**
Peter Valentin-Weigand (Hannover)
- 10:15 Coffee break
- 10:45 **Insights into influenza virus infections with and without bacteria**
Thorsten Wolff (Berlin)
- 11:30 **Subversion of inflammatory responses by the pneumococcus**
Jeffrey N. Weiser (New York, USA)
- 12:30 Lunch

Session II: Biosafety (Workshop)

Venue: **Alfried Krupp Wissenschaftskolleg Greifswald**

Chair: **Linda Brunotte & Thorsten Wolff**

- 13:30 **Animal biosafety**
Jens Teifke (Island of Riems)
- 14:00 **Biosafety – Risk assessment on research of viral-bacterial co-infections**
Jürgen Mertsching (Hannover)
- 14:30 Discussion
- 15:00 Coffee break and poster session in the entrance foyer

Program

Session III: Ethics and Dual use issues in the Biological Sciences (Panel Discussion)

Venue: Alfried Krupp Wissenschaftskolleg Greifswald

Chair: Christian Suhm & Sven Hammerschmidt

- 17:00 Thomas C. Mettenleiter (Greifswald - Insel Riems)
Bärbel Friedrich (Greifswald)
Jörg Hacker (Berlin)
Lothar Wieler (Berlin)
Hans-Dieter Klenk (Marburg)
Linda Brunotte (Muenster)
- 19:00 Dinner

Friday, September 30th, 2016

Session IV: Host-Pathogen-Interactions of Bacteria and Viruses

Venue: Alfried Krupp Wissenschaftskolleg Greifswald

Chair: Hans-Dieter Klenk & Peter Valentin-Weigand

- 09:00 [Insights into the role of the transcription termination factor Rho of *Staphylococcus aureus*](#)
Anna Nagel (Greifswald)
- 09:20 [The hidden lipoproteome of *Staphylococcus aureus*](#)
Anica Beyer (Greifswald)
- 09:40 [Bat influenza virus chimeras as basis for the development of a new type of vaccine backbone for livestock vaccination](#)
Jacob Schön (Greifswald - Insel Riems)
- 10:00 [Proteomic analysis of lymphocytes after infection and transformation with the oncogenic Gallid herpes virus 2](#)
Viktoria Pauker (Greifswald - Insel Riems)
- 10:20 Coffee break and poster session in the entrance foyer
- 12:00 Lunch

Program

Session V: Imaging and OMICS technologies in the Infection Research (Workshop)

Venue: Friedrich-Loeffler-Institut

Chair: Katharina Riedel & Jan Pané-Farré

- 14:00 [Exploration of viral life cycles by fluorescent virus imaging](#)
Stefan Finke (Greifswald - Insel Riems)
- 14:30 [Molecular pathogen-host interaction in human lung tissue](#)
Andreas Hocke (Berlin)
- 15:00 Discussion
- 15:45 Coffee break
- 16:15 [Proteomics and Immunoproteomics - lessons learned from *S. aureus* host-pathogen interactions](#)
Frank Schmidt (Greifswald)
- 16:45 [Detection and characterization of pathogens by high-throughput sequencing](#)
Dirk Höper (Greifswald - Insel Riems)
- 17:15 Discussion
- 18:00 [Farewell](#)
Sven Hammerschmidt (Greifswald)
Thomas C. Mettenleiter (Greifswald - Insel Riems)
- 18:30 Barbecue

Venues

Alfried Krupp Wissenschaftskolleg Greifswald

Martin-Luther-Straße 14

17489 Greifswald

Germany

Friedrich-Loeffler-Institut

Suedufer 10

17493 Greifswald - Insel Riems

Germany



Subversion of Inflammatory Responses by the Pneumococcus

Weiser JN

Department of Microbiology, New York University School of Medicine, New York, USA

Much of the mortality attributed to influenza virus is due to secondary bacterial pneumonia, particularly from *Streptococcus pneumoniae*. We find that prior influenza infection enhances the two critical steps in pneumococcal pathogenesis; colonization of the nasopharynx and host-to-host transmission.

Influenza accelerates bacterial replication *in vivo*, and sialic acid, a major component of airway glycoconjugates, is identified as the host-derived metabolite that stimulates pneumococcal proliferation. Influenza infection increases sialic acid and sialylated mucin availability and enhances desialylation of host glycoconjugates. Pneumococcal genes for sialic acid catabolism are required for influenza to promote bacterial growth. Decreasing sialic acid availability *in vivo* by genetic deletion of the major airway mucin *Muc5ac* or mucolytic treatment limits influenza-induced pneumococcal replication.

We adapted an infant mouse model to elucidate host determinants of transmission of *S. pneumoniae* from inoculated index mice to uninfected contact mice. In the context of co-infection with influenza A virus, the pneumococcus is transmitted among wildtype littermates. Mice deficient for TLR2 are colonized to a similar density but transmit *S. pneumoniae* more efficiently than wildtype animals and show a decrease in expression of interferon α and higher viral titers. The greater viral burden in *tlr2*^{-/-} mice correlates with heightened inflammation, and is responsible for an increase in bacterial shedding from the mouse nose. Taken together, these results suggest that the innate immune response to influenza virus promotes bacterial shedding, allowing the bacteria to transit from host to host.

Nakamura, S, KM Davis and JN Weiser. Synergistic stimulation of type I interferons during influenza co-infection promotes *Streptococcus pneumoniae* colonization in mice. *Journal of Clinical Investigation*. 121(9):3657-65. 2011

Siegel, SJ, and JN Weiser. Influenza promotes pneumococcal growth during co-infection by providing host sialylated substrates as a nutrient source. *Cell Host & Microbe*. 16:55-67. 2014

Richard, AL, SJ Siegel, J Erikson, and JN Weiser. TLR2 signaling decreases transmission of *Streptococcus pneumoniae* by limiting bacterial shedding in an infant mouse influenza A co-infection model. *PLoS Pathogens*. 10:e1004339. 2014

Contact: Jeffrey N. Weiser
Jeffrey.Weiser@nyumc.org



Animal Biosafety

Teifke J

Department of Experimental Animal Facilities and Biorisk Management, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

The number of institutions that keep a variety of animal species for research purposes on infectious, especially zoonotic diseases is growing worldwide. The same is true for public awareness and concerns about occupational health, environmental protection, and last but not less important, animal welfare and for the body of regulations on these issues. All these fields are covered by the term "Animal Biosafety". When working with infected animals, especially livestock species, specific care must be taken to prevent the spread of disease. Thus, in-vivo research requires each investigator and all laboratory personnel to take additional safety precautions in the primary containment of large animal rooms to minimize the incidence of laboratory acquired infections. To ensure a safe environment for those who work with infected animals it is crucial that the occupational and environmental health and safety program of an institution is drawn up in close cooperation with the Institutional Animal Care and Use Committee (IACUC) and an Institutional Biorisk Committee (IBC) which coordinates and surveys all issues of biosafety and biosecurity. This places significant responsibility on employers to ensure both the appropriate level of animal care and protection of the employees but also of agriculture involved in animal research.

This presentation provides information on the use of experimentally infected animals housed in the high containment research facilities at the Friedrich-Loeffler-Institut (FLI) with a focus on settings where these animals harbor and propagate zoonotic pathogens or infectious agents of high veterinary significance for their use in translational research.

1. Fontes B. Institutional responsibilities in contamination control in research animals and occupational health and safety for animal handlers. *ILAR J.* 2008;49(3):326-37.
2. Abad, FX; Solanes, D; Domingo, M. Animal Biosafety Level 3 facility - enhancements when dealing with large animals. In: *Working in Biosafety Level 3 and 4 laboratories : a practical introduction / ed. by Manfred Weidmann, Nigel Silman, Patrick Butaye and Mandy Elschner. - Weinheim : Wiley Blackwell, 2014: 31-39.*
3. Frasier D, Talka J. Facility design considerations for select agent animal research. *ILAR J.* 2005;46(1):23-33.

Contact: [Jens P. Teifke](mailto:JensPeter.Teifke@fli.de)
JensPeter.Teifke@fli.de

Insights into the role of the transcription termination factor Rho of *Staphylococcus aureus*

Nagel A,¹ Nicolas P,² Debarbouille M,³ Michalik S,¹ Hertlein T,⁴ Depke M,¹ Guerin C,² Hiron A,³ van der Kooi-Pol MM,⁵ Palma Medina LM,¹ Murr A,¹ Pané-Farré J,⁶ Hecker M,⁶ Ohlsen K,⁴ Msadek T,³ van Dijl JM,⁵ Völker U,¹ Mäder U¹

¹Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald, Germany

²MalAGE, INRA, Université Paris-Saclay, Jouy-en-Josas, France

³Department of Microbiology, Institut Pasteur, Paris, France

⁴Institute for Molecular Infection Biology, University of Würzburg, Germany

⁵Department of Medical Microbiology, University of Groningen, University Medical Center Groningen, The Netherlands

⁶Institute for Microbiology, University of Greifswald, Germany

In a comparative OMICs-study we analyzed *S. aureus* HG001, a derivative of the strain NCTC 8325, under multiple experimental conditions. Data of the tiling array transcriptome analysis [1] revealed a relatively low abundance of antisense RNAs in the *S. aureus* wild type, where they overlap only 6% of the coding genes. As known from previous studies, the transcription termination factor Rho plays a major role in suppressing antisense transcription in *E. coli* [2] and *B. subtilis* [3], and indeed there is a remarkable overall increase in antisense transcription in the absence of Rho in *S. aureus*. In contrast to results reported for *E. coli*, elevated antisense transcription significantly affected sense transcript levels. Proteome analysis of cytoplasmic and secreted fractions showed significant differences in the abundance of several proteins, namely increased amounts of SaeSR-dependent virulence factors like extracellular adherence protein (Eap) and fibronectin-binding proteins (FnbA and FnbB) in the rho mutant. Elevated expression of the regulon controlled by the SaeSR two-component system perfectly confirmed the mRNA data of the tiling array study. Our data suggest that under conditions of Rho deficiency the SaeSR regulatory system of *S. aureus* is activated by a so far unknown mechanism leading to the observed induction of the SaeSR regulon. Currently, we use different infection models to further explore the impact of the higher levels of SaeSR-dependent virulence factors.

[1] Mäder et al.; PLOS Genet. 2016; 12(4):e1005962.

[2] Peters et al.; Genes Dev. 2012; 26(23):2621-33.

[3] Nicolas et al.; Science 2012; 335(6072):1103-6.

Contact: Anna Nagel
anna.nagel@uni-greifswald.de



Bat influenza virus chimeras as basis for the development of a new type of vaccine backbone for livestock vaccination

Schön J,¹ Hoffmann D,¹ Juozapaitis M,² Wei R,² Schwemmler M,² Beer M¹

¹Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald – Insel Riems, Germany

²Institute of Virology, University Medical Center Freiburg, Freiburg, Germany

Influenza-A-viruses (IAVs) are important pathogens for a broad range of animals like birds, horses, dogs and pigs. Beside seasonal infections in humans leading to annual epidemics, IAV-spillover infections from animals to humans can result in severe infections with a huge economically impact. Moreover, livestock and companion animals are acting as mixing vessels for different IAVs, bearing the risk that reassortants are able to cross the species barrier. The most recent 2009 influenza pandemic in humans is e.g. based on an avian-swine-human quadruple reassortant virus. Effective and safe IAV live vaccines for animals do not exist with very few exceptions for pigs outside of Europe. This is mainly due to the risk of possible reassortment events between vaccine strains and wild type IAV-strains, and partly because of the pathogenicity associated with live vaccine strains especially in very young animals. However, we recently succeeded to generate chimeric viruses containing six out of the eight bat influenza A-like H17N10 virus genes, with the remaining two genes encoding the haemagglutinin and neuraminidase proteins of a prototypic IAV. These viruses grow in mammalian and avian cells, but exhibited limited replication in chicken. Most importantly, these bat chimeras fail to reassort with other IAV. Therefore, bat influenza A-like viruses are to our opinion a very promising vector backbone for the development of a new generation of live IAV vaccines. Determining and characterizing different chimeric combinations with improved properties for live vaccines using in vitro and in vivo settings are our objects of research.

Contact: [Jacob Schön](mailto:jacob.schoen@fli.de)
jacob.schoen@fli.de

Molecular pathogen-host interaction in human lung tissue

Hocke AC,¹ Zscheppang K,¹ Berg J,¹ Faykhova D,¹ Nerlich A,¹ Mieth M,¹ Opitz B,¹ Becher A,¹ Szymanski K,¹ Rückert, JC,² Neudecker J,² Tönnies M,³ Bauer TT,³ Eggeling S,⁵ Schimek M,⁵ Wolff T,⁴ Suttrop N,¹ Hippenstiel S¹

¹Department of Internal Medicine/Infectious Diseases and Respiratory Medicine, Charité – Universitätsmedizin Berlin, Germany

²Department of General, Visceral, Vascular and Thoracic Surgery, Charité – Universitätsmedizin Berlin, Germany

³Helios Clinic Emil von Behring, Department of Pneumology and Department of Thoracic Surgery, Berlin, Germany

⁴Division of Influenza and Other Respiratory Viruses, Robert Koch-Institute, Berlin, Germany

⁵Vivantes Clinic Neukölln, Department of Thoracic Surgery, Berlin, Germany

Pneumonia is a leading cause of death worldwide and mortality rates are constant since decades. *Streptococcus pneumoniae* (*S. pneumoniae*) is the most common bacterial respiratory pathogen causing community-acquired pneumonia and preceding influenza infections regularly pave the way for even more severe bacterial infections. The pathogens directly induce the host immune system, which involves cellular damage and death. However, the underlying mechanisms differ between pathogens, species, and get complex in viral-bacterial co-infections.

Therefore, we established a human lung tissue model by which viral and bacterial infections alone as well as in combination are analysed by e.g. molecular, biochemical, or microscopic procedures. Next to lab strains, the model serves to investigate clinically isolated pathogens, which allows for comparison of different virulence factors.

We found a unique cellular tropism of different influenza subtypes for type II pneumocytes and showed that these cells also contribute to the inflammatory response against *S. pneumoniae*. Whereas influenza cause a non-inflammatory apoptosis of these cells during its replication, pneumococci cause a pneumolysin dependent lytic cell death showing signs of apoptosis and necrosis. Moreover, pneumococcal pneumolysin induce IL-1beta in alveolar macrophages leading to protective epithelial GM-CSF release. In contrast, influenza triggers a strong interferon response interfering with pneumococcal activation of the IL-1/GM-CSF-axis, whereas cell death induction remained unaffected.

The use of living human lung tissue excellently serves for the investigation of basic molecular mechanisms determining the development of infectious diseases. Thereby, it reveals differences to mice with regard to the IL-1/GM-CSF-axis and is useful to test pharmacological intervention.

1. Influenza A viruses target type II pneumocytes in the human lung. Weinheimer VK1, Becher A, Tönnies M, Holland G, Knepper J, Bauer TT, Schneider P, Neudecker J, Rückert JC, Szymanski K, Temmesfeld-Wollbrueck B, Gruber AD, Bannert N, Suttrop N, Hippenstiel S, Wolff T, Hocke AC. J Infect Dis. 2012
2. Streptococcus pneumoniae-induced regulation of cyclooxygenase-2 in human lung tissue. Szymanski KV1, Toennies M, Becher A, Fatykhova D, N'Guessan PD, Gutbier B, Klauschen F, Neuschaefer-Rube F, Schneider P, Rueckert J, Neudecker J, Bauer TT, Dalhoff K, Drömann D, Gruber AD, Kershaw O, Temmesfeld-Wollbrueck B, Suttrop N, Hippenstiel S, Hocke AC. Eur Respir J. 2012
3. Influenza induced interference with pneumococcal host immune activation. Berg J, Zscheppang K, Fatykhova D, Toennies M, Schneider P, Rueckert J, Neudecker J, Bauer TT, Eggeling S, Schimek M, Gruber AD, Kershaw O, Suttrop N, Hippenstiel S, Hocke AC. In submission

Contact: [Andreas Hocke](mailto:andreas.hocke@charite.de)
andreas.hocke@charite.de

Platelet activation and aggregation provoked by Staphylococcus aureus secreted proteins

Binsker U,¹ Wesche J,² Palankar R,² Kohler TP,¹ Prucha J,¹ Bröker B,³ Mamat U,⁴ Pané-Farré J,⁵ Beyer A,⁵ Schmidt F,⁶ Greinacher A,² Hammerschmidt S¹

¹Department Genetics of Microorganisms, Interfaculty Institute for Genetics and Functional Genomics, University of Greifswald, Germany

²Department of Transfusion Medicine, Institute of Immunology and Transfusion Medicine, University Medicine Greifswald, Germany

³Department of Immunology, Institute of Immunology and Transfusion Medicine, University Medicine Greifswald, Germany

⁴Division of Structural Biochemistry, Research Center Borstel, Leibniz-Center for Medicine and Biosciences Borstel, Germany

⁵Department of Microbial Physiology and Molecular Biology, Institute for Microbiology, University of Greifswald, Germany

⁶Department of Functional Genomics, Interfaculty Institute for Genetics and Functional Genomics, University of Greifswald, Germany

Staphylococcus aureus is an opportunistic human pathogen provoking a wide range of severe community-acquired and nosocomial infections amongst others infective endocarditis (IE) and disseminated intravascular coagulopathy (DIC). Both clinical manifestations constitute an uncontrolled activation of both platelets and coagulation cascade resulting in thrombocytopenia. S. aureus - platelet interactions occur either directly or indirectly via recruitment of serum components. Bacterial factors inducing platelet activation or aggregation are mostly secreted proteins possessing ECM binding activity.

Fifty-six recombinant secreted or surface-localized staphylococcal proteins were screened for their capacity to activate platelets, measured by the activation markers P-selectin and α IIb β 3 conformation, using whole blood, platelet-rich-plasma (PRP), and washed platelets in buffer from a constant set of donors. Micropattern protein array (MiPA) chips were functionalized with His6-tagged staphylococcal proteins to assess the interactions on a single cell level.

This study confirmed the potential of the SERAM (secretable expanded repertoire adhesive molecules) protein Eap to induce platelet activation and aggregation. In addition, this study further identified the chemotaxis inhibitory protein CHIPS, the formyl peptide receptor-like 1 inhibitory protein FLIPr, all involved in immune evasion, as well as the major autolysin Atl as potent platelet activators. Furthermore, the domains of Atl and the extracellular adherence protein (Eap), responsible for platelet activation could be narrowed down.

Taken together, this study identified two members of the SERAM family (Eap and Atl) and two additionally secreted proteins of S. aureus as platelet activators and aggregators. These results emphasize the importance and diversity of S. aureus-platelet interactions.

Contact: Ulrike Binsker
binskeru@uni-greifswald.de



Staphylococcus aureus lipase 1 – a novel staphylococcal immunomodulatory factor?

Dick J,¹ Kolata J,² Zehrfeld C,¹ Behrendt A,¹ Bornscheuer U,³ Bröker B¹

¹Department of Immunology, University Medicine Greifswald, Germany

²Department of Medical Microbiology – University Medical Center Utrecht, Netherlands

³Department of Biotechnology & Enzyme Catalysis – Institute of Biochemistry – Ernst Moritz Arndt University Greifswald, Germany

Staphylococcus aureus is a common commensal but can also cause severe infections. This ambiguity is usually explained by the multifaceted interactions between the bacterium and the immune system. S. aureus produces various virulence factors that interfere with different parts of the immune system. Still, the human immune system is usually able to keep the bacterium at bay, and various studies show that a partially protective immunological memory can be established. For instance, B cells produce antibodies against most extracellular staphylococcal proteins. However, there is one protein standing out: The Staphylococcus aureus lipase 1 (Lip), a secreted lipolytic enzyme which is highly abundant and very conserved, elicits only extremely low levels of specific IgG and IgM antibodies in humans. The goal of this project is to unravel the reasons for the observed antibody gap.

First results indicate that Lip has a strong mitogenic effect on T lymphocytes. Stimulation with active Lip induced very strong proliferation of T cells, suggesting polyclonal T cell activation. This effect was abolished when using a nonfunctional S408A mutant. In this case, the proliferation indices as well as Lip-specific precursor frequencies were comparable to those of typical S. aureus recall antigens.

The T cell activating effect therefore appears to be dependent on the enzymatic function, while typical staphylococcal superantigens rely on their structure.

The next steps towards a better understanding of Lip's function will involve a closer look at the prerequisites for proliferation induction as well as the functional characterization of the proliferating T cells.

Contact: Johannes Dick
johannes.dick@uni-greifswald.de

Simple, quick and cost-efficient: A universal RT-PCR for amplifying and sequencing the genome of foot-and-mouth disease virus.

Dill V, Beer M, Hoffmann B

Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Foot- and mouth disease (FMD) comes along with debilitations and restrictions for livestock industries all over the world. Because of its high contagiousity prevention strategies are of great importance. The foot-and-mouth disease virus (FMDV) is included in the family of Picornaviridae, genus Aphthovirus, possessing a small single-stranded RNA genome of 8.4 kilobases (kb). Generation of whole genome sequences is of increasing importance due to the growth of international nucleotide databases. It provides epidemiological tracing of virus transmission and is supportive in case of virus disease control strategies. This study describes the development and examination of a quick, universal and cost-efficient RT-PCR assay to generate the sequence of the whole polyprotein of the FMDV genome. The novel RT-PCR method was evaluated using twelve different virus strains from cell culture, covering all seven serotypes of FMDV. Additionally, different sample materials were tested to mimic diagnostic or rather field samples. All primer pairs showed a robust amplification with high sensitivity and with no evidence of a better performance for Eurasian serotypes than for SAT serotypes. In summary, the described assay is suitable for the generation of FMDV sequences covering the whole polyprotein to be used in phylogenetic analysis, quick serotyping and antigenic characterisation.

Contact: Veronika Dill
veronika.dill@fli.de

Cell adhesion molecules (integrins) modulate Flavivirus infection in mouse cell lines

dos Reis VP,¹ Keller M,¹ Schmidt K,² Groschup MH¹

¹Institute for Novel and Emerging Infectious Diseases, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Deutsches Krebsforschungszentrum, Mikrobiologische Diagnostik, Heidelberg, Germany

To infect the host cells, viruses must interact with a broad range of molecules which includes their cellular receptors and molecules that support virus replication. Flaviviruses like Dengue, West Nile, Usutu, Yellow fever and Zika virus have caused recent outbreaks around the world. Up to now, few molecules were characterized as Flavivirus receptor(s) or able to modulate flavivirus infection. Integrins are a family of transmembrane proteins expressed in many cell types with important functions like cell migration, attachment, mitoses and apoptosis. Previous studies postulated that West Nile virus uses integrin to enter into the cells and further studies from our group showed that integrins modulate West Nile virus infection. In order to elucidate if integrins are involved in Flavivirus infection we infected integrin deficient mouse fibroblasts and CHO cells, a flavivirus resistant cell line, with different Flaviviruses. In these sets of experiments we evaluated: (i) virus binding; (ii) replication kinetics; (iii) internalization and (iv) replication. Our results show that integrins are not the main flavivirus receptor and its expression in CHO cells did not confer Flavivirus permissiveness. Also, integrins are not involved in Flavivirus binding neither internalization into the cell. Replication was slightly impaired in cells deficient for $\beta 1$ and $\beta 3$ integrin subunits and strongly impaired in $\alpha V\beta 3$ integrin deficient cells. In summary, integrins are not involved in the first steps of Flavivirus infection (binding and internalization) but somehow integrins modulate Flavivirus replication. The exact mechanism how integrins modulate flavivirus replication is unknown and subject of investigation.

Contact: Vinicius Pinho dos Reis
Vinicius.Reis@fli.de



Comparative pathogenicity study of European bat lyssavirus 1 isolates in the mouse model

Eggerbauer E,¹ Finke S,¹ Mettenleiter TC,¹ Müller T,¹ Nolden T,² Pfaff F,³ Teifke J,⁴ Freuling CM¹

¹Institute of Molecular Virology and Cell Biology, WHO Collaborating Centre for Rabies Surveillance and Research, Friedrich-Loeffler-Institut, Greifswald – Insel Riems, Germany

²ViraTherapeutics GmbH, Innsbruck, Austria

³Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

⁴Department of Experimental Animal Facilities and Biorisk Management, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

European bat lyssavirus 1 (EBLV-1) is the most common of the four lyssavirus species present in the bat population in Europe and known to have caused rabies in a variety of mammals including humans. The genome of EBLV-1 isolates displays a certain heterogeneity, visible in their division into two genetic distinct sublineages as well as in insertions and deletions which were found in recent years. The impact of these genetic differences on the pathogenicity of EBLV-1 has never been assessed even though it was observed for other lyssaviruses, i.e. Lagos bat virus. Therefore we compared the pathogenicity of eight EBLV-1 isolates, representing both sublineages as well as known insertions and deletions, in the mouse model using different routes of inoculation. Besides survival, we also investigated the amount of viral antigen present in different brain areas of mice which succumbed to rabies. Results show that differences in the pathogenicity of the isolates exist when inoculated intranasally or intramuscularly with a low dose. Furthermore variations in the incubation periods could be observed following intramuscular inoculation. Clinical symptoms as well as the viral loads in different parts of the brain depend on the inoculation route. We therefore conclude that results obtained with one EBLV-1 isolate cannot be generalized for the whole species and studies using only single isolates as representatives need careful interpretation.

Contact: [Elisa Eggerbauer](mailto:Elisa.Eggerbauer@fli.de)
Elisa.Eggerbauer@fli.de



Establishment of serological assays for the detection of henipavirus specific antibodies in pigs

Fischer K,¹ dos Reis VP,¹ Diederich S,¹ Groschup MH,¹ Weingartl HM,² Balkema-Buschmann A¹

¹Institute of Novel and Emerging Infectious Diseases, Friedrich-Loeffler-Institut, Greifswald-Insel Riems, Germany

²Canadian Food Inspection Agency, National Centre for Foreign Animal Disease, Winnipeg, Canada

Hendra and Nipah virus (HeV; NiV) are enveloped, single-stranded negative-sense RNA viruses in the Henipavirus genus, Paramyxoviridae family. Henipavirus infections were first reported in the 1990's causing severe and often fatal outbreaks in domestic animals and humans in the Australasian region. Recently, there has been first serological evidence for the presence of henipa-like viruses in African fruit bats, pigs and humans.

Truncated forms of henipavirus attachment (G) proteins were expressed in a novel expression system based on the stable transfection of the eukaryotic parasite *Leishmania tarentolae*. Both G proteins were applied in Enzyme-linked Immunosorbent Assay (ELISA) for the detection of henipavirus specific antibodies in serum samples of pigs experimentally infected with HeV or NiV.

HeV and NiV G proteins were efficiently purified via Strep-tag affinity chromatography with total protein yields of about 1 mg per litre of cell culture. For NiV challenged pigs, seroconversion was detectable 14 days after infection in the NiV G based ELISA whereas the HeV G ELISA only detected cross-reactive henipavirus specific antibodies at day 20 post infection. Sera of HeV infected pigs collected 28 days after infection cross-reacted with the NiV G but, as expected, displayed higher reactivity in the HeV G based assay.

Finally, the henipavirus G proteins expressed in our study were successfully used to test henipavirus challenged pig sera in ELISA. Serological testing of field sera from African pigs against both *Leishmania*-derived henipavirus G proteins will present a valuable approach to indirectly determine the presence of henipa-like virus specific antibodies.

Contact: Kerstin Fischer
Kerstin.Fischer@fli.de



Survey for the zoonotic bacteria *Leptospira* and *Rickettsia* in wild small mammal populations in Germany

Fischer S,¹ Mayer-Scholl A,² Essbauer S,³ Kratzmann N,¹ Imholt C,⁴ Reil D,⁴ Heuser E,¹ Schmidt S,¹ Rosenfeld UM,¹ Jacob J,⁴ Nöckler K,² Ulrich RG¹

¹Institute for Novel and Emerging Infectious Diseases, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Federal Institute for Risk Assessment (BfR), Department for Biological Safety, Berlin, Germany

³Bundeswehr Institute of Microbiology, Munich, Germany

⁴Julius Kühn-Institute (JKI), Institute for Plant Protection in Horticulture and Forests, Münster, Germany

Amongst wildlife species, rodents are considered to be an important reservoir for various zoonotic bacteria. Here, we describe a *Leptospira* and *Rickettsia* survey of 4,019 investigated rodents and shrews in Germany, performed between 2010 and 2014 in rural settings in Mecklenburg-Western Pomerania, North Rhine-Westphalia, Thuringia and Baden-Wuerttemberg.

Kidney tissue samples were analyzed by a PCR targeting the *lipL32* gene. Using this initial screening PCR, 524 of 3,950 (13.3%) small mammals were tested positive for *Leptospira*. Subsequently, a partial *secY* gene-specific PCR and sequencing were used to identify the *Leptospira* species. The presences of *Leptospira kirschneri*, *Leptospira interrogans* or *Leptospira borgpetersenii* were determined. MLST-based typing resulted in the identification of six sequence types. Spotted Fever Group *Rickettsia*-specific DNA was detected in 314 of 3,939 (8.0%) ear pinna samples, using a real-time PCR targeting the citrate synthase (*gltA*) gene. Typing of the rickettsial positive species was performed by a conventional PCR, targeting the partial outer membrane protein B (*ompB*) gene. The *ompB*-based typing resulted in the identification of three *Rickettsia* species, *Rickettsia helvetica*, *Rickettsia felis* and *Rickettsia raoultii*. In conclusion, leptospiral and rickettsial DNA could be found in each rodent and small mammal species with a broad geographical distribution, but without distinct host specificity. Co-infections of *Leptospira* and *Rickettsia* were found at low frequency of 0.8%. These findings underline the need for further investigations to assess the public health relevance of the different potential animal reservoirs and pathogen lineages.

Contact: Stefan Fischer
stefan.fischer@fli.de



Metabolic and regulatory adaption of *Staphylococcus aureus* to different carbon sources

Giese A,¹ Dörries K,² Zühlke D,¹ Liang C,³ Lalk M,² Hecker M,¹ Riedel K,¹ Dandekar T,³ Pané-Farré J¹

¹Institute of Microbiology, Ernst-Moritz-Arndt-Universität Greifswald, Greifswald, Germany

²Institute of Biochemistry, Ernst-Moritz-Arndt-Universität Greifswald, Greifswald, Germany

³Department of Bioinformatics, Biocenter, Julius-Maximilians-Universität Würzburg, Würzburg, Germany

Different human body sites are characterized by specific carbon sources available to *S. aureus*. For example, during invasive disease glucose is the major carbon source in the blood stream, whereas pyruvate may correlate with a colonizing lifestyle since it is available in significant amounts in nasal secretion. Lactate for instance is found in lungs of cystic fibrosis patients where it is secreted by *Pseudomonas aeruginosa* during co-colonization with *S. aureus*. In addition, lactate may also function as a signal of the hosts' state of immune defense since lactate is produced in significant amounts by proliferating T-cells at the site of infection. Finally, for many intracellular pathogens glycerol has been demonstrated as a major carbon source during host cell invasion and thus may have an equally important role for *S. aureus* during intracellular survival which is an important step in the development of chronic *S. aureus* infections.

For a better understanding of *S. aureus* adaptation processes to different host-environment relevant carbon sources we are using a combined metabolomic and proteomic approach. *S. aureus* was grown in a synthetic medium with glucose, pyruvate, lactate or glycerol as sole carbon source. To also investigate the impact of carbon catabolite repression in this process, we included a catabolite control protein A mutant in our analyses and exposed *S. aureus* to combinations of glucose with either pyruvate, lactate or glycerol. Quantitative data will be presented describing the protein repertoire and extracellular metabolic profile of *S. aureus* when grown with different carbon sources.

Contact: Anne Giese
anne.giese@uni-greifswald.de

EVAg – European Virus Archive goes global: a unique biological resource in the field of virology

Goller KV,¹ Pabion M,² Prat C,² Romette JL,² Teifke JP,¹ Mettenleiter TC,¹ Reiche S¹

¹Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Aix-Marseille Université, Marseille, France

During the past century and especially the last decade with upcoming new technologies, a large number of pathogenic viruses with emerging potential have been discovered and isolated. These viruses are routinely used for basic research, epidemiological studies, disease control, development of diagnostics as well as for therapeutic purposes like the generation of novel vaccines and antivirals. Therefore, besides appropriate skills and capacities, the availability of suitable high quality reference material is crucial. In 2009, a FP7 project called "European Virus Archive" (EVA) was conceived whereby the overall objective was to constitute a European network of high calibre centres with the appropriate expertise to identify, collect, amplify, characterise, standardise, authenticate, and distribute viruses. EVA has been operating satisfactorily and about 2000 products have been distributed worldwide. In order to continue and expand its activities the H2020 project "European Virus Archive goes Global" (EVAg; <http://www.european-virus-archive.com/>) has been founded by the European Commission, and consist of an international group of 25 laboratories. The ultimate goal of this non-profit organisation is to become the largest virus collection worldwide and provide eligible access to valuable high quality guaranteed viruses of all risk groups as well as virus-derived materials like viral genomes and proteins, suitable cell lines, antigen-specific sera or monoclonal antibodies, and to unique services including access to high biosafety level laboratories and animal facilities. Accordingly, EVAg is mainly involved in the development and distribution of diagnostic reference material e.g. during the emergence of viruses such as MERS-CoV or Zika virus.

Contact: Katja Goller
katja.goller@fli.de



Pneumococcal fitness and virulence gene regulation by the TCS08

Gómez Mejía A,¹ Petruschka L,¹ Gámez G,² Böhm S,² Kluger V,² Klein A,² Mäder U,³ Hammer-schmidt S¹

¹Department Genetics of Microorganisms, Interfaculty Institute for Genetics and Functional Genomics, University of Greifswald, Greifswald, Germany

²Basic and Applied Microbiology (MICROBA) Research Group, School of Microbiology, Universidad de Antioquia, Medellín, Colombia

³Department of Functional Genomics, Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald, Greifswald, Germany

Background: *Streptococcus pneumoniae* (pneumococci) is equipped with virulence factors that are coordinated by various regulatory systems. Among these systems, the Two-Component regulatory Systems (TCS) play a major role in pneumococcal fitness and virulence. Hence, the impact of TCS08 and its effect on the regulation of virulence and metabolism in *S. pneumoniae* was evaluated in this study. Methods: To assess the impact of the TCS08 in pneumococci, isogenic mutants (Δ rr08, Δ hk08 and Δ tcs08) were generated. MBP-tagged recombinant HK08 and RR08 were purified by affinity chromatography. Functional assays (phostag-acrylamide, electromobility shift assay (EMSA), Transcriptomics) and a pneumonia model were performed. Results: A substantially increased expression of the PavB protein was detected only for the hk08-mutant. The phosphotransfer assays with TCS08 proteins suggests autophosphorylation of RR08 and a phosphatase activity of HK08. The EMSA with purified RR08 and a pavB promoter DNA-fragment illustrated binding of non- and phosphorylated RR08. The microarray analysis showed regulation of different genes involved in virulence and metabolism. Finally, in vivo infections in the acute pneumonia mouse model displayed a lower virulence capacity for the Δ hk08 and Δ tcs08 mutants but not Δ rr08. Conclusion: These data show that TCS08 regulates genes involved in pneumococcal pathogenicity and metabolism. Additionally this study suggests that non-phosphorylated RR08 functions as a repressor while the phosphorylated RR08 acts as an activator for the expression of PavB. Finally, Δ hk08 and Δ tcs08 mutants had a lower virulence and were unable to progress post-intranasal infection from the lungs to the blood in mice.

Contact: Alejandro Gómez Mejía
alejandro.gomezmejia@uni-greifswald.de



Entry and reassortment potential of the newly discovered bat influenza A-like viruses

Gorka M,¹ Hoffmann D,¹ Stertz S,² Schwemmler M,³ Beer M¹

¹Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Institute of Medical Virology, University of Zürich, Switzerland

³Institute for Virology, University Medical Center Freiburg, Germany

Influenza A viruses (IAV) are important zoonotic pathogens that cause epidemic outbreaks in poultry, wild birds, swine and other mammals. In 2012 and 2013 two bat influenza A-like virus genomes were found in little yellow-shouldered fruit bats (*Sturnira lilium*) in Guatemala and flat-faced fruit bats (*Artibeus planirostris*) in Peru, provisionally designated as H17N10 and H18N11. Infectious virus particles, however, were never isolated and attempts to generate infectious viruses by reverse genetic approaches failed so far. Furthermore conventional IAV hemagglutinins (HAs) bind canonical sialic acid-containing receptors. In contrast, biochemical and structural studies indicated that influenza A-like H17 does not. In fact, H17 and H18 HAs are unable to bind and hemagglutinate red blood cells, and are therefore atypical HAs. Besides, the neuraminidase (NA) works as a counterpart to HA in the life cycle of conventional IAV. It is an enzyme that cleaves sialic acids from their sugar backbone to facilitate the release of newly synthesized viral particles from infected cells. Bat influenza A-like NAs however lack any detectable neuraminidase activity. These two differences lead to the suggestion, that bat influenza A-like viruses use other entry mechanisms, compared to conventional IAV. With a first series of experiments we aim to identify the functional entry receptor. We also hypothesize that NA-deleted classical IAVs are perfect surrogate viruses to study either bat NA or bat HA or both bat proteins. To achieve this, bat HA and the NA will be integrated in NA-deleted H5 and H7 viruses by a reverse genetics approach.

Contact: Marco Gorka
Marco.Gorka@fli.de

A natural pair of low and high pathogenicity avian influenza virus H7N7 in Germany

Graaf A,¹ Beer M,¹ Mettenleiter T,² Harder T¹

¹Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald – Insel Riems, Germany

²Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald – Insel Riems, Germany

Avian influenza viruses threaten poultry production worldwide. Highly pathogenic avian influenza viruses (HPAIV) cause devastating losses in gallinaceous poultry and, due to zoonotic propensities, also pose a threat to public health. The last outbreak of HPAIV H7N7 in Germany (one holding in Emsland) dates back to summer 2015. This virus likely arose by spontaneous mutation from a low pathogenic (LP) H7N7 precursor virus detected in a neighbouring chicken holding. It remains to be elucidated how, when and why such mutations occur. The availability of this native LP/HP pair of viruses enables further work into the mechanisms of HPAIV generation in a project that started in April 2016:

- The presence of HP mutations in LPAIV H5 and H7 quasispecies from field samples will be examined by RT-qPCR and NGS.
- Competing co-infections of LP and HP AIV H7N7/Emsland will be carried out in chickens and ducks to determine the minimal percentage of HP viruses in a mixture required to induce disease.
- Generation of HP AIV H7N7/Emsland from its LP precursor by serial passages of LP virus in eggs and cell cultures of different avian species and in vivo will be attempted in order to gain insights into mutation mechanisms.

Contact: [Annika Graaf](mailto:Annika.Graaf@fli.de)
Annika.Graaf@fli.de

Arenavirus Life Cycle Modelling Systems: Development and Application

Leske A,¹ Dunham EC,² Shifflett K,² Watt A,² Hoenen T,^{2,3} Groseth A^{1,2}

¹Independent Research Group Arenavirus Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Laboratory of Virology, Division of Intramural Research, National Institute of Allergy and Infectious Diseases (NIAID), National Institutes of Health (NIH), Hamilton, MT, USA

³Institute for Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Arenaviruses are responsible for a number of viral hemorrhagic fever (VHF) diseases endemic to Africa and South America. These viruses are transmitted from rodents to humans via aerosols, particularly those contaminated with rodent excreta, and in some cases possibly also ingestion of contaminated food. VHF-causing arenaviruses are classified as biosafety level (BSL) 4 agents, which poses significant challenges for both research and the development of antivirals. Our goal was to develop a variety of systems that can be safely used under BSL1/2 conditions for both basic and applied research applications.

To this end minigenomes (genome analogues that replace the viral proteins with reporter proteins such as GFP or Luciferase) were constructed for Junin virus. These constructs were then expressed together with the viral polymerase complex proteins (NP and L) in mammalian cells to facilitate viral transcription/replication, resulting in reporter protein synthesis, which serves as a read-out for these processes. Further, to expand this system, the viral proteins responsible for budding and entry (GP and Z) were co-expressed along with the minigenome components, leading to the generation of transcription and replication-competent virus-like particles (trVLPs), which can then be used to infect target cells.

Using these assays we have conducted analyses looking at aspects of basic virus biology (e.g. transcription/replication efficiency as a pathogenic determinant), performed applied screens for potential antiviral compounds and host factors important for the viral lifecycle, and most recently adapted these assays for the detection of neutralizing antibodies.

Contact: Allison Groseth
allison.groseth@fli.de

Interaction of Matrix Proteins with Nuclear ANP32B Is Conserved Among Henipa- and Avulaviruses

Günther M, Bauer A, Zaack L, Römer-Oberdörfer A, Finke S

Institute for Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Recently, the nuclear cell factor acidic leucine-rich nuclear phosphoprotein 32 family member B (ANP32B) has been identified as a nuclear interactor of Hendra and Nipah virus (HeV, NiV; genus Henipavirus) matrix proteins. Although the detailed mechanism still has to be resolved, the specificity of this interaction and re-localisation of the matrix proteins (M) strongly suggested that ANP32B plays an important role in host manipulation by henipaviruses.

In order to see, whether interaction with ANP32B represents a conserved feature among paramyxoviruses, we here investigated whether M proteins of Newcastle Disease Virus (NDV, genus Avulavirus, subfamily Paramyxovirinae), Bovine Respiratory Syncytial Virus (BRSV, genus Pneumovirus, subfamily Pneumovirinae) and Measles virus (MV, genus Morbillivirus, subfamily Paramyxovirinae) also interact with ANP32B. Notably, ANP32B was successfully co-purified with NDV M and, similar to henipavirus M proteins, NDV M accumulated in the nucleus in an ANP32B dependent manner. In contrast, no co-purification of BRSV M and MV M with ANP32B was observed, indicating that both does not interact with ANP32B. Our data strongly support the hypothesis, that ANP32B is part of a conserved host cell manipulatory mechanism of Henipa- and Avulaviruses. Ongoing mutational analysis are performed to identify conserved binding motifs and to further unravel the mechanism behind.

Contact: Maria Günther
maria.guenther@fli.de

Responses of bovine lung epithelial cells to *Mycoplasma mycoides* strains, which have different target hosts

Hänske J,¹ Golchert J,² Homuth G,² Heller M,³ Jores J,⁴ Kammerer R¹

¹Institute of Immunology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald, Greifswald, Germany

³Institute of Molecular Pathogenesis, Friedrich-Loeffler-Institut, Jena, Germany

⁴Institute of Veterinary Bacteriology, University of Bern, Bern, Switzerland

Contagious bovine pleuropneumonia (CBPP) caused by *Mycoplasma mycoides* subsp. *mycoides* (Mmm) is an important cattle disease in Africa. The present control measures rely on attenuated live vaccines with limited efficiency and sporadic severe side effects at the site of inoculation. The knowledge of host pathogen interactions and the protective immune mechanisms will foster the development of an improved vaccine against CBPP.

We investigated the very first host response by analyzing the transcriptome of embryonic bovine lung cells (EBL), which were co-cultivated with the highly virulent African strain Afadé (Mmm) and that of EBL cells co-cultivated with the caprine strain *Mycoplasma mycoides* subsp. *capri* (Mmc), that is non-pathogenic for cattle but the closest relative of Mmm. Co-cultured with Mmm EBL showed a reduction of cell viability. Particularly noteworthy is the upregulation of transcripts encoding interferon induced proteins, like ISG15, IFIT1 and MX1. Even Mmc led to a remarkable upregulation of genes, confirming a strong impact of this pathogen to bovine cells. However this response did not affect the viability of the eukaryotic cells. Surprisingly, the caprine mycoplasma adhered better to bovine cells than the bovine pathogen. Nevertheless, in contrast to EBL in co-cultivation with the caprine strain, EBL co-cultivated with Mmm were not able to create a protective response against the pathogen leading to cell death. Although there is a common response at both strains, the inappropriate response to Mmm led to a deleterious reaction that was not observed with Mmc, characterizing the pathogenic mechanisms of Mmm.

Contact: Jana Hänske
jana.haenske@fli.de



Identification of a pneumococcal enzyme essential for anchoring of lipoteichoic acid to the bacterial cell surface

Heß N,¹ Waldow F,² Kohler T,¹ Gisch N,² Hammerschmidt S¹

¹Dept. Genetics of Microorganisms, Interfaculty Institute for Genetics and Functional Genomics, Uni Greifswald

²Division of Bioanalytical Chemistry, Priority Area Infections, Research Center Borstel, Leibniz-Center for Medicine and Biosciences

Typical structures of Gram-positive cell walls are wall teichoic acids (WTA) and lipoteichoic acids (LTA) which are highly diverse and often species- and strain-specific. The role of WTA in cell morphology and division, regulation of autolysis and adhesion to host cells is e.g. well characterized for *Staphylococcus aureus* and *Bacillus subtilis*, while the role of LTA remains elusive. Teichoic acids (TA) of *Streptococcus pneumoniae* are unique in several aspects. First, pneumococcal WTA (pnWTA) and LTA (pnLTA) are identical in their complex repeating unit structures, suggesting a common biosynthesis pathway. Second, pnTA are highly decorated with phosphorylcholine (P-Cho), which bind non-covalently choline-binding proteins (CBPs) to the surface and are essential for bacterial growth. Based on the *in silico* analysis of the *S. pneumoniae* R6 genome, putative enzymes involved in the biosynthetic pathway of teichoic acids have been proposed. Inactivation of *spd_1672*, a gene in strain D39 encoding an enzyme with homology to O-antigen ligases from different Gram-negative bacteria, demonstrated the involvement of this enzyme in pnLTA anchoring to the surface of pneumococci. Chemical analysis of the cell wall of mutant D39Δ*cps*Δ*spd1672* indicated a total loss of pnLTA compared to the isogenic wild-type. The phenotypic characterization of the mutant revealed an impact on growth and an increased autolytic activity. Alterations in cell morphology of pneumococcal wild-type and mutants were illustrated by SEM. Furthermore, quantification of selected TA associated CBPs and capsule was investigated by flow cytometry. In summary, we present here the identification of an enzyme involved in anchoring of pnLTA.

Contact: Nathalie Heß
hessn@uni-greifswald.de



Network "Rat-borne pathogens": Searching for pathogen co-infections

Heuser E,¹ Fischer S,¹ Ryll R,¹ Mayer-Scholl A,² Hoffmann D,³ Spahr C,² Imholt C,⁴ Johne R,² Ehlers B,⁵ Essbauer S,⁶ Nöckler K,² Ulrich RG^{1,7}

¹Institute for Novel and Emerging Infectious Diseases, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Federal Institute for Risk Assessment, Berlin, Germany

³Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald-Insel Riems, Germany

⁴Julius Kühn-Institute, Federal Research Centre for Cultivated Plants, Institute for Plant Protection in Horticulture and Forestry, Vertebrate Research, Münster, Germany

⁵Robert Koch-Institute, Berlin, Germany

⁶Bundeswehr Institute of Microbiology, Munich, Germany

⁷German Center for Infection Research (DZIF), partner site Hamburg-Luebeck-Borstel-Insel Riems, Germany

The Norway rat *Rattus norvegicus* is an important reservoir of zoonotic pathogens, such as orthopox virus (OPV) and *Leptospira*, but also for agents of no or unknown zoonotic potential. In addition, in Norway rats human pathogens have been detected, but rats are most likely not involved in their transmission, but may serve as a sentinel.

In a recent survey, Norway rats originating from five European countries were investigated for *Leptospira* spp., *Rickettsia* spp., OPV, and rat polyomavirus (ratPyV). *Leptospira* DNA was detected in 60 of 420 rats and *Rickettsia* DNA was found in three of 369 rats. PCR-based typing resulted in the identification of *L. interrogans* and *Rickettsia helvetica*, respectively. RatPyV DNA was detected in 103 of 421 rats. OPV DNA was detected in none of the rats, but OPV-specific antibodies in three of 388 rats. The frequency of single *Leptospira* and ratPyV infections and co-infections was, independent of sex, greater for adults compared to juveniles/subadults and greater at rural sites compared to urban areas.

Study results indicate a broad geographical distribution of *Leptospira* DNA in rats within Europe underlining the need to further investigate potential mechanisms leading to increased prevalence in rural habitats. In contrast, rickettsia and OPV infections rarely occurred in wild rat populations. The potential influence of ratPyV on the susceptibility to infections with other pathogens should be investigated in future studies. Recently, pilot studies were initiated within the network for *Staphylococcus aureus*, *Streptobacillus moniliformis* and further pathogens.

Contact: [Elisa Heuser](mailto:Elisa.Heuser@fli.de)
Elisa.Heuser@fli.de



Investigations of the phosphoproteome of *Streptococcus pneumoniae*

Hirschfeld C,¹ Hoyer J,¹ Hentschker C,¹ Junker S,¹ Gomez A,² Maab S,¹ Hammerschmidt S,² Becher D¹

¹Institute for Microbiology, University Greifswald, Greifswald, Germany

²Interfaculty Institute for Genetics and Functional Genomics, University of Greifswald, Greifswald, Germany

Streptococcus pneumoniae is a human pathogen that can cause several severe and invasive infections. Due to the worldwide rise of resistance to antibiotics in pneumococci it is of great relevance to analyze not only the pneumococcal genome but also the proteome. Data about protein abundance in *S. pneumoniae* may provide an extensive source of information for the selection of new targets for vaccine and antibiotic development.

Protein phosphorylation is a reversible post-translational modification enabling the translation of extracellular signals into cellular responses and adaptation to a steadily changing environment. Therefore, it is of particular interest to capture precise qualitative and quantitative information about the phosphoproteome of *S. pneumoniae*.

S. pneumoniae has only one gene encoding a PP2C-type phosphatase, PhpP, located upstream from the *stkP* gene, which encodes the only membrane-associated serine-threonine kinase, StkP. PhpP and StkP are in the focus of our study. They appear to constitute a functional signaling couple *in vivo* and likely belong to the same complex.

For the identification and quantification of relative differential changes in the phosphoproteome of the unencapsulated wildtype (WT) and the deletion mutants Δ stkP and Δ phpP we use the metabolic labeling strategy Stable isotope labeling using amino acids in cell culture (SILAC) followed by the enrichment of phosphorylated peptides with TiO₂ and mass spectrometric analysis.

First results show, that the cell division proteins FtsZ and DivIVA are phosphorylated in the WT. Furthermore we found phosphorylation sites in some ABC-transporters and cation transporters that might play a role in various biological processes.

Contact: Claudia Hirschfeld
claudia.hirschfeld@uni-greifswald.de

Investigations to *Streptococcus pneumoniae* under iron limitation

Hoyer J,¹ Hirschfeld C,¹ Hentschker C,¹ Gomez A,² Hammerschmidt S,² Maaß S,¹ Becher D¹

¹Institute for Microbiology, University Greifswald, Greifswald, Germany

²Interfaculty Institute for Genetics and Functional Genomics, University Greifswald, Greifswald, Germany

Streptococcus pneumoniae is one of the major human pathogens, which can cause several severe diseases ranging from mild forms like sinusitis to life-threatening infections like septicemia. They can lead to high mortality and morbidity rates, especially among children under the age of five, elderly and immunosuppressed patients. Therefore it is important to understand how the pneumococcal virulence functions, not only on genome but also on proteome level.

The acquisition of iron is essential for pathogenic bacteria. Iron is involved in several key metabolic pathways both as a cofactor and redox-active catalyst. Because of the high insolubility of iron under physiological conditions, the iron concentration in its human host is restricted. Hence, the aim of this study is to analyze the changes of the proteome of *S. pneumoniae* under iron limitation compared to control conditions.

In order to quantify changes in protein abundances we used SILAC as a metabolic labeling technique. Furthermore, we used 2,2'-bipyridine to induce iron limitation. After cultivation, the sample were fractionated in cellular and extracellular proteins and pooled with a heavy labeled internal SILAC standard prepared under the conditions examined. Followed by LC-MS/MS analysis, the obtained data were subjected to a classical database search and statistically analyzed. In this study more than 800 of 1914 predicted pneumococcal proteins were identified. 500 proteins could be quantified, including about 130 significantly regulated proteins. A part of those proteins belong to pathogenesis- and iron-associated proteins, like the pneumococcal surface protein C, PspC, and the iron-compound ABC transporters SPD_0915 and SPD_1652.

Contact: [Juliane Hoyer](mailto:hoyerj@uni-greifswald.de)
hoyerj@uni-greifswald.de



The Newcastle disease virus W protein – an mRNA editing product of the phosphoprotein gene

Karsunke J,¹ Karger A,¹ Franzke F,² Hammerschmidt B,³ Mettenleiter TC,¹ Römer-Oberdörfer A¹

¹Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Institute of Infectology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

³Department of Experimental Animal Facilities and Biorisk Management, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Like all members of the Paramyxovirinae Newcastle disease virus (NDV) undergoes a co-transcriptional mRNA editing to enlarge its coding capacity. The P gene codes for a 395 amino acids (aa) phosphoprotein. By co-transcriptional insertion of one or two non-templated G nucleotides at the RNA editing site two further mRNAs are transcribed, coding for V and W proteins. The 293 aa V protein is a known interferon antagonist. However, the deduced W protein of 176 aa has so far not been demonstrated.

Here, the existence of W protein was investigated for NDV Clone 30, a lentogenic vaccine virus strain. Mass spectrometry investigations of purified virions gave the first indication for the presence of the W protein. For further confirmation, W specific synthetic peptides were selected for immunization of rabbits to generate W protein specific serum. The obtained serum was then used for indirect immunofluorescence, Western blot analyses and localization studies by electron and confocal microscopy, confirming the existence of the W protein in cell lysates as well as in purified virions.

In summary, we show for the first time the existence of the NDV W protein.

Contact: [Julia Karsunke](mailto:Julia.Karsunke@fli.de)
Julia.Karsunke@fli.de

Detection of novel viral pathogens by adaptive diagnostics to prevent epidemics

Kauer L,¹ Höper D,¹ Jenckel M,¹ Belka A,¹ PeiBert C,² Lembcke R,² Reimer U,³ Knaute T,³ Holenya P,³ Eckey M,³ Noack K,² Beer M,¹ Pohlmann A¹

¹Institute of Diagnostic Virology, Friedrich-Loeffler-Institute, Greifswald – Insel Riems, Germany

²Scopeland Technology GmbH, Berlin, Germany

³JPT Peptide Technologies GmbH, Berlin, Germany

The basis for research in infection biology is the detection and characterization of the responsible pathogen, e.g. viruses. Since there are no diagnostic tools for novel viruses they often remain undetected. Infectious diseases which cause no or only mild symptoms in their host, but lead to severe symptoms after a change of species, are particularly dangerous. Next-generation sequencing (NGS) combined with a metagenomic analysis provides an opportunity to detect and characterize new viruses. However, it provides no information about the serological host response and the development of serological assays is often complex and time-consuming. Since there is a gap between the detection of new viruses and necessary serological diagnostics, the BMBF-funded interdisciplinary project DetektiVir aims to develop a generic workflow that is applicable to all kinds of novel virus diseases. The workflow starts with a sample containing an unidentified virus, in which via NGS and metagenomic analysis viral sequences can be identified and classified. These identified viral sequences are the basis for the subsequent identification of suitable antigenic regions – by using peptide array analysis – finally resulting in a functional serological assay. Furthermore, the proposed workflow lays the foundation for future research on a detected novel virus in the field of infection biology. The first part of the workflow is now already optimized and will be presented with a few examples highlighting the strengths and weaknesses of the concept.

Contact: [Leonie Kauer](mailto:leonie.kauer@fli.de)
leonie.kauer@fli.de

B-cell response of mice against RVFV strain MP12

König R, Keller M, Eiden M, Rissmann M, Groschup MH

Institute of Novel and Emerging Infectious Diseases, Friedrich-Loeffler-Institute, Greifswald – Insel Riems, Germany

Rift Valley Fever Virus is a zoonotic virus transmitted by mosquitos. Main hosts are ruminants but mice are found highly susceptible too. MP12 is an attenuated strain of RVFV used for vaccination of cattle and other domestic ruminants. Mice immunized with MP12 should show high antibody titers with neutralizing activity in VNT.

In former studies at the FLI RVFV strain MP12 was used in mice to generate monoclonal antibodies. Therefore, mice received several boosts after immunization and the spleens were used for the generation of monoclonals. In contrast to these studies the time line of the B-cell response based on seroconversion following a one-shot immunization of mice is in focus in this study.

After immunization serum was examined in regular time lag for antibodies against the glycoproteins Gn and Gc as well as the nucleoprotein NP. After seroconversion was confirmed a serum neutralization assay with MP12 was performed to determine if the antibodies are protective against reinfection. To further characterize the immune response to envelope proteins an ELISA based pespcan method was used to determine binding patterns to the glycoproteins Gn and Gc.

With this study the exact time point for the appearance of protective virus neutralizing antibodies can be determined. It may serve as a model for testing of other RVFV vaccines in mice.

Contact: [Rebecca König](mailto:rebecca.koenig@fli.de)
rebecca.koenig@fli.de

Mapping domains of the Schmallenberg virus nonstructural protein NSm

Kraatz F, Wernike K, Aebischer A, Reimann I, Beer M

Institute of Diagnostic Virology, Friedrich-Loeffler-Institute, Greifswald – Insel Riems, Germany

Schmallenberg virus (SBV) is an insect-transmitted orthobunyavirus causing severe fetal malformation, abortions and stillbirth in ruminants. While the nonstructural protein NSs is a major virulence factor, the main functions of NSm are still unknown.

To investigate the role of the integral membrane-protein NSm, containing three hydrophobic transmembrane (I, III and V) and two nonhydrophobic regions (II and IV), we generated a number of recombinants on the basis of our reverse genetics system and tested these constructs in virus rescue experiments, growth kinetics and interferon-assays.

Rescue experiments were successful with mutants containing a complete deletion of NSm domain IV, or a partial C-terminal deletion within domain II or when both deletions were combined. Mutants with partial N-terminal deletions of domain II and complete deletion of domain III, respectively, could not be rescued. All replicating single-mutants as well as NSm and NSs double-mutants displayed only mild differences in their growth properties in BHK-21 cells. Interferon-induction was only observed with double-mutants with a NSs deletion.

In conclusion, our data show that the N-terminal parts of the NSm domains II and III are essential for the generation of infectious virus, whereas domain IV and the C-terminal part of domain II are not required.

Contact: Franziska Kraatz
Franziska.Kraatz@fli.de

Development of a theophylline-based riboswitches for gene regulation in two obligate intracellular pathogens *Coxiella burnetii* and *Chlamydia abortus*

Lämmer F, Anders V, Steinert J, Berens C

AG260 Molecular Pathogenetic, Institute of Molecular Pathogenesis, Friedrich-Loeffler-Institut, Jena, Germany

Bacterial gene expression is frequently controlled at the RNA level by regulatory elements called riboswitches. They consist of an aptamer sequence, which recognizes a specific ligand and an effector domain which controls gene expression at the transcriptional or translational level via an RNA conformational switch. Natural riboswitches bind metabolites like coenzyme B12 to regulate the respective biosynthetic pathway. Additionally, artificial riboswitches employing synthetic ligands like theophylline active in many organisms. They are attractive tools because their one component nature makes them self-sufficient, easy to implement and respond rapidly. Therefore, I want to introduce riboswitch regulation to the obligate intracellular pathogens *Coxiella burnetii* and *Chlamydia abortus* using a set of published theophylline-responsive riboswitches, shown to work in different organisms. The constructs use RFP as reporter gene, different plasmid backgrounds and promoter sequences in *Escherichia coli* as initial model. First, I tested the strong Tac promotor, active in all three organisms. Later, it will be replaced by organism specific promoters like the com1 promoter from *C. burnetii* or the ompA promoter from *C. abortus*. Different RSF1010-based vectors serve as shuttle plasmids for both organisms. First experiments show that the riboswitches function in *E. coli* and that they differ in their regulatory properties.

Contact: Frauke Laemmer
frauke.laemmer@fli.de

Development of Novel Double-Attenuated Live Vaccines in Swine against H1N1 Influenza Strains

Mamerow S, Stech O, Scheffter R, Mettenleiter TC, Stech J

Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Pigs are commercially relevant livestock and frequently infected with influenza viruses. The disease is of great economic relevance and furthermore bears high zoonotic risks. To reduce disease burden and minimize virus reservoirs, effective vaccination is still a key issue. However, conventional inactivated vaccines often confer insufficient protection and are strongly biased towards a predominant humoral immune response. For that reason, this project aims to improve this situation in pig husbandry by developing a live attenuated influenza virus vaccine. By reverse genetics, we generated attenuated mutants of the wild-type strain A/Bayern/74/2009 (H1N1v). We cloned all eight gene segments and introduced a non-physiological, strictly elastase-dependent cleavage site into the hemagglutinin. For this purpose, we replaced the amino acids at positions four to one (P4-P1) upstream of the actual proteolytic cleavage site by four alanine residues. In addition, we have truncated the C terminus of the non-structural protein 1 to generate a double-attenuated mutant. After we had studied the in-vitro properties, we performed the first in-vivo experiment to investigate the attenuation features of the single-attenuated virus mutant Bayern74/09/HA-Ela as well as the double-attenuated virus mutant Bayern74/09/HA-Ela/NS1-99.

Contact: [Svenja Mamerow](mailto:svnja.mamerow@fli.de)
svnja.mamerow@fli.de

Toxoplasma gondii: Characterization of virulence factors from genetically atypical cross products

Matzkeit B,¹ Herrmann DC,¹ Höper D,² Maksimov P,¹ Knittler M,³ Conraths FJ,¹ Schares G¹

¹Institute of Epidemiology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Institute of Diagnostic Virology, Friedrich-Loeffler-Institute, Greifswald - Insel Riems, Germany

³Institute of Immunology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

The obligate intracellular protozoan parasite *Toxoplasma gondii* has a clonal population structure in Europe and North America. Among these clonal types only *T. gondii* strains of type I are highly virulent for immunocompetent lab mice, e.g. BALB/c. In Germany, the less virulent *T. gondii* type II strains prevail. The variance in mouse virulence can be explained by strain dependent polymorphisms of parasitic virulence factors that can interact differentially with the host immune system. *T. gondii* oocysts representing type II/III recombinant genotype were isolated from a fecal sample obtained from a naturally infected cat in Germany. From this isolate five individual parasite clones were generated as tachyzoites, each showing different combinations of type II and III allele patterns. The clones also revealed differences in virulence for BALB/c mice. The known prominent parasite virulence factors ROP18 and ROP5 that are able to inhibit the activity of mouse immunity related GTPases (IRG), were analyzed to clarify the distinct virulence behavior in BALB/c mice. The ROP18 expression differs markedly whereas the ROP5 allele is the same in the 5 clones. Furthermore, the mRNA expression profile of the cytokines IL-12, IL-10, and TNF α , which activity can be influenced by the *T. gondii* virulence factors GRA15 and ROP16, was determined in IFN γ activated macrophages. The results show that the parasites differ in their ability to stimulate the cytokine expression.

Contact: Beate Matzkeit
Beate.Matzkeit@fli.de



Generation of recombinant Newcastle disease virus expressing Peste des petits ruminants virus surface glycoproteins

Murr M,¹ Hoffmann B,² Mettenleiter TC,¹ Römer-Oberdörfer A¹

¹Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Peste des petits ruminants (PPR) is a highly contagious disease in small ruminants as sheep and goats caused by the Peste des petits ruminants virus (PPRV) belonging to the genus morbillivirus in the family paramyxoviridae.

The surface glycoproteins hemagglutinin (H) and fusion protein (F) are considered to be the most important viral proteins to induce a protective immune response in the host since neutralizing antibodies are being generated towards H and F during infection.

In order to differentiate between infected and vaccinated animals, recombinant marker vaccines (DIVA vaccines) are useful. Therefore, we generated two recombinant Newcastle disease viruses (NDV) expressing either the PPRV H or F protein by reverse genetics. The expression of the PPRV glycoproteins was confirmed by western blot analysis of infected cells.

The two recombinant viruses replicate well in embryonated SPF chicken eggs, their replication in different sheep and goat cell lines as well as their thermostability was investigated.

In summary, two recombinant NDV are available which can be studied in an animal experiment for their fitness to protect from PPR, a disease that becomes more important because of the eradication of rinderpest and the abolition of vaccination against it which conveyed a cross protection against PPRV.

Contact: Magdalena Murr
magdalena.murr@fli.de

Cell Culture Adaptation of Non-fixed Rabies Virus Strains from Fox and Dog

Nemitz S,¹ Christen M,² Pfaff F,³ Finke S¹

¹Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Ernst-Moritz-Arndt University, Greifswald, Germany

³Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Fixed Rabies virus (RABV) laboratory strains, like SAD, are generally known to infect worse via peripheral routes of inoculation compared to non-fixed isolates. Vice versa, non-fixed field isolates grow bad on standard cell cultures and need some passages to adapt. Starting with recombinant virus clones from non-adapted dog and fox field viruses (rRABV DogA, rRABV Fox 148), we performed serial passages on murine neuroblastoma cells (NA), canine epithelial cells (MDCK) and hamster kidney cells (BsrT7/5) and monitored infectious virus titers in cell culture supernatants. Since titers are increased by more than 2 logs while passaged on BsrT7/5 cells, deep-sequencing after the 10th passage was performed to show cell culture adaptation. Whereas most viral genes remained conserved, mutations accumulated in the glycoprotein. In fox and dog virus identical amino acid exchanges were identified. In addition to an amino acid exchange at position 266 present in all passaged viruses, a BsrT7/5 cell specific mutation at position 444 was selected, indicating that cell type or species specific constraints exist that restrict cell culture replication of field RABV. Confocal fluorescence microscopy further indicated that the identified mutations caused less extensive ER retention of G and more efficient virus particle release at the plasma membrane.

Contact: Sabine Nemitz
sabine.nemitz@fli.de



Rupture Forces among Human Blood Platelets at different Degrees of Activation

Nguyen TH, Palankar R, Bui VC, Medvedev N, Delcea M, Greinacher A

Institute for Immunology and Transfusion Medicine, University Medicine Greifswald, Greifswald, Germany

Little is known about mechanics underlying the interaction among platelets during activation and aggregation. Although the strength of a blood thrombus has likely major biological importance, no previous study has measured directly the adhesion forces of single platelet-platelet interaction at different activation states. Here, we filled this void first, by minimizing surface mediated platelet-activation and second, by generating a strong adhesion force between a single platelet and an AFM cantilever, preventing early platelet detachment. We applied our setup to measure rupture forces between two platelets using different platelet activation states, and blockade of platelet receptors. The rupture force was found to increase proportionally to the degree of platelet activation, but reduced with blockade of specific platelet receptors. Quantification of single platelet-platelet interaction provides major perspectives for testing and improving biocompatibility of new materials; quantifying the effect of drugs on platelet function; and assessing the mechanical characteristics of acquired/inherited platelet defects.

Contact: Thi-Huong Nguyen
nguyent@uni-greifswald.de

Allergic reactions to Staphylococcal serine proteases in airway diseases

Nordengrün M,¹ Teufelberger A,² Michalik S,^{3,4} Stentzel S,¹ Schmidt F,^{3,4} Krysko O,² Völker U,³ Bachert C,^{2,5} Bröker BM¹

¹Department of Immunology, University Medicine Greifswald, Germany

²Upper Airways Research Laboratory, Ghent University, Belgium

³Interfaculty Institute for Genetics and Functional Genomics, Department Functional Genomics, University Medicine Greifswald, Germany

⁴Junior Group Applied Proteomics, ZIK FunGene, University Medicine Greifswald, Germany

⁵Division of Ear, Nose and Throat Diseases, Clintec, Karolinska Institute, Stockholm, Sweden

Recently there is increasing evidence that besides commensal and invasive behavior *S. aureus* may also drive allergic reactions. In patients suffering from allergic disorders, e.g. atopic dermatitis and nasal polyposis, *S. aureus* colonization appears much more frequent. Additionally, allergic airway diseases are frequently associated with high titer IgE binding to staphylococcal superantigens. An implication of *S. aureus* proteins in the pathophysiology of allergic disorders has thus been suggested. However, the driving allergens of *S. aureus* remained elusive.

We could identify the serine protease-like proteins (Spl) A-F as major IgG4-binding proteins of *S. aureus*. Spl-specific IgG1 and IgG4 was quantified in sera of healthy donors (n=46) by ELISA. Spl-specific IgE was measured in serum of asthma patients (n=50) and healthy controls (n=40). Peripheral blood T cells of healthy donors (n=9) were stimulated with Spls and cytokine production was elucidated by cytometric bead array. The ability of Spls to induce allergic responses in vivo was tested in a murine asthma model.

Compared to Hla, the Spl-specific antibody response was significantly shifted toward IgG4. Spl-specific IgE was significantly elevated in serum of asthma patients. Spl-stimulation of T cells elicited a Th2/Treg-dominated cytokine profile (IL-4, IL-5, IL-10, IL-13), whereas Th1/Th17 cytokines (IFN- γ , IL-6, IL-17, TNF- α) were of low concentrations or absent. In mice, inhalation of SplD without adjuvant induced allergic lung inflammation characterized by Th2 cytokines and eosinophil infiltration.

The results indicate a role of *S. aureus* Spls as triggering allergens in allergic airway diseases, opening prospects for diagnosis and causal therapy of asthma.

Contact: Maria Nordengrün
maria.nordengruen@uni-greifswald.de



Immunome analysis reveals species-specific humoral immune response in sepsis patients

Normann N,¹ Tietz G,¹ Stentzel S,¹ Gerber M,² Guderian L,² Gründling M,² Balau V,³ Nauck M,⁴ Steinmetz I,³ Schulz K,³ Völker U,⁵ Meissner K,² Bröker BM,¹ Kolata J^{1,6}

¹Department of Immunology, University Medicine Greifswald, Germany

²Department of Anaesthesiology and Intensive Care Medicine, University Medicine Greifswald, Germany

³Friedrich Loeffler Institute of Medical Microbiology, University Medicine Greifswald, Germany

⁴Institute of Clinical Chemistry and Laboratory Medicine, University Medicine Greifswald, Germany

⁵Department of Functional Genomics, University Medicine Greifswald, Germany

⁶Current position: Medical Microbiology, University Medical Center Utrecht, The Netherlands

Sepsis is the third leading cause of death in hospitalized patients in Germany. Blood culture is positive in less than half of the cases of clinical sepsis. Yet the identification of the causative agent is essential for a targeted antimicrobial therapy.

In this study we examined whether the ability of sepsis patients to generate a highly specific humoral immune response towards the invading pathogen might support conventional sepsis diagnosis.

In a prospective clinical trial ICU patients with suspected sepsis were recruited. Plasma samples were collected before sepsis, at diagnosis and during the infection. Plasma antibody binding to extracellular proteins of frequent sepsis pathogens was quantified using a Simple Western Assay. Dynamics in antibody binding were assessed in 74 patients. Healthy adults served as control. 2D-immunoblots were used to identify immunogenic proteins of different bacteria.

In healthy subjects, anti-bacterial IgG levels were stable over at least two months. In contrast, sepsis patients frequently exhibited dynamic antibody profiles during sepsis. An increase of specific IgG could be detected in 40% of patients with a microbiological diagnosis (21/53) and also in one third of sepsis patients without a suspected pathogen (7/21). In cases with an intraabdominal infection focus, an antibody response to more than one bacterial species was often observed.

During sepsis the adaptive immune system responds specifically to the invading pathogen. The identification of immunogenic proteins of frequent sepsis pathogens may help to develop a new serological test for pathogen detection and risk stratification in sepsis patients.

Contact: Nicole Normann
nicole.normann@uni-greifswald.de



Analysis of vaccine-induced rabies cases using deep sequencing

Pfaff F,¹ Freuling C,² Müller T,² Beer M,¹ Mettenleiter TC,² Höper D¹

¹Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Rabies is a long known zoonotic disease caused by an ssRNA(-) virus of the genus Lyssavirus. During large-scale vaccination programs of foxes, millions of oral rabies vaccine virus baits were distributed. Notably, most of them contained one of the live-attenuated SAD-derived (Street Alabama Dufferin) strains B19 or P5/88, both in vitro derivatives of SAD-Bern(orig). Post vaccination surveillance detected at least ten possible vaccine-associated fatal rabies cases in four different species: five in red foxes (*Vulpes vulpes*) and for each one in stone marten (*Martes foina*), European badger (*Meles meles*), and cattle (*Bos taurus*), respectively. It was shown that genetic relationships of SAD vaccines based on consensus sequence information could lead to biased results and the important role of low frequency variants was highlighted. In order to determine the genetic relationship of the selected vaccine-associated viruses with the vaccine strains used for the vaccination campaign in the respective region, we conducted deep sequencing. For data analysis beyond consensus distances we used the frequency of each nucleotide at each position of the viral genome to calculate pairwise Manhattan distances from a nucleotide frequency alignment of the sequenced strains. Results were visualized using two-dimensional non-metric distance scaling. Isolates from vaccine-induced rabies cases clustered tightly together, but surprisingly none of the SAD-B19-related cases grouped around their parental vaccine batches. Instead these isolates showed similarities to other in vitro derivatives of SAD-Bern(orig) indicating an in vivo selection process favoring only a portion of the variants/haplotypes within SAD-B19 vaccines.

Contact: Florian Pfaff
Florian.Pfaff@fli.de

Molecular Requirements for Nuclear Egress of Herpesviruses

Propp S, Hellberg T, Klupp BG, Mettenleiter TC

Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Herpesviruses use a vesicle-mediated process for translocation of newly formed capsids from the nucleus to the cytoplasm where final virus maturation proceeds. For this, capsids bud at the inner nuclear membrane, acquiring a primary envelope which is subsequently lost after fusion with the outer nuclear membrane (ONM) thereby releasing the capsids into the cytoplasm. This vesicle-mediated translocation engaging the nuclear membranes was long considered as unique for herpesviruses but recent data suggest a similar, yet not well characterized regular cellular pathway.

For herpesviruses vesicle budding and scission is solely orchestrated by the nuclear egress complex consisting of the two viral proteins designated as pUL31 and pUL34 in the alphaherpesviruses. This was shown not only in eukaryotic cells but also in model membrane systems indicating that no other viral or cellular protein is required for the formation of primary enveloped virions located in the perinuclear space. However, it is still completely unknown which viral and/or cellular proteins mediate the release of the capsids to the cytosol orchestrating the fusion of the primary virion envelope with the ONM

To test which proteins might be involved, we will isolate primary enveloped virions from the perinuclear space. This is not an easy task but we already established cell lines stably expressing a tagged pUL34 to use this in affinity chromatography. The enriched primary enveloped virions will be then analyzed by mass spectrometry to identify the putative fusion machinery in the nuclear envelope which might also mediate nuclear translocation of cellular components.

Contact: [Sebastian Propp](mailto:sebastian.propp@fli.de)
sebastian.propp@fli.de

Validation of Gene Candidates Obtained by Comparative Prediction

Romoth L, Gerischer L, König S, Stanke M

Institute of Mathematics and Computer Science

Many hundreds of strains have recently been sequenced for many bacterial species. This provides an opportunity to improve gene finding by exploiting this variation as well as it poses the challenge of consistent annotation of these strains.

An extension of the gene finder Augustus allows the simultaneous prediction of genes in several aligned bacteria genomes. This comparative version decides for gene candidates by a score that is based on the one from single-genome Augustus, that is based on a conditional random field, but also incorporates evidence for selection. It rewards a consistent annotation of aligned regions using a phylogenetic tree of the genomes. A dual-decomposition algorithm is used to approximate the maximal score [König 2015].

We applied comparative Augustus to all strains with a complete assembly on NCBI of each of the 5 bacterial species *Staphylococcus aureus*, *Streptococcus pneumoniae*, *Burkholderia pseudomallei*, *Bacillus subtilis* and *Escherichia coli*. To this end the genomes were aligned with Cactus [Paten 2011] for every species. As input for Cactus we passed nearly star-shaped trees. We found that the genes that are unique to the Augustus gene set have a higher percentage of homologs in the species specific NCBI protein databases than the genes that are unique to the RefSeq annotations. Further, we used other information like occurrence of a recognisable ribosome binding site motif, gene length and quantitative RNA data (if available) as indicator for correctly predicted genes.

Contact: [Lars Romoth](mailto:lars.romoth@uni-greifwald.de)
lars.romoth@uni-greifwald.de

Rat hepatitis E virus in Europe: Looking for viral and bacterial co-infections

Ryll R,¹ Bernstein S,¹ Heuser E,¹ Schlegel M,^{1,17} Dremsek P,^{1,18} Zumpe M,^{1,19} Wolf S,^{2,20} Pépin M,³ Bajomi D,⁴ Müller G,⁵ Heiberg AC,⁶ Spahr C,⁷ Lang J,⁸ Groschup MH,¹ Ansorge H,⁹ Freise J,¹⁰ Guenther S,¹¹ Imholt C,¹² Heckel G,^{13,14} John R,¹⁵ Ulrich RG,^{1,16}

¹Institute for Novel and Emerging Infectious Diseases, Friedrich-Loeffler-Institut, Greifswald-Insel Riems, Germany

²Institute for Microbiology, Technische Universität Dresden, Dresden, Germany

³VetagroSup (Microbiologie/Immunologie/Pathologie infectieuse), Coordinateur de l'Unité d'Enseignement de MEDECINE PREVENTIVE, USC INRA-VAS 1233 / Equipe PERS („Pathogènes Emergents et Rongeurs Sauvages“), Marcy l'étoile, France

⁴Bábolna Bio Ltd., Budapest, Hungary

⁵Stadt Zürich, Umwelt- und Gesundheitsschutz Zürich, Zürich, Switzerland

⁶AC Heiberg Rådgivning, Mørkøv, Denmark

⁷German Federal Institute for Risk Assessment, Berlin, Germany

⁸Institut für Tierökologie und Naturbildung, Gonterskirchen, Germany

⁹Senckenberg Museum of Natural History, Görlitz, Germany

¹⁰Niedersächsisches Landesamt für Verbraucherschutz und Lebensmittelsicherheit, Fachbereich Schädlingsbekämpfung, Task-Force Veterinärwesen, Wardenburg, Germany

¹¹Institute of Microbiology and Epizootics, Veterinary Faculty, Freie Universität Berlin, Berlin, Germany

¹²Julius Kühn-Institute, Federal Research Centre for Cultivated Plants, Institute for Plant Protection in Horticulture and Forestry, Vertebrate Research, Münster, Germany

¹³University of Bern, Institute of Ecology and Evolution, Bern, Switzerland

¹⁴Swiss Institute of Bioinformatics, Quartier Sorge - Batiment Genopode, Lausanne, Switzerland

¹⁵German Federal Institute for Risk Assessment, Berlin, Germany

¹⁶German Center for Infection Research (DZIF)

¹⁷Seramun Diagnostica GmbH, Heidesee, Germany

¹⁸Labor Doz. DDr. Stefan Mustafa, Wien, Austria

¹⁹University Medicine Greifswald, Pediatric Hematology and Oncology, Ferdinand-Sauerbruch-Straße, Greifswald, Germany

²⁰Labor Ostsachsen, Dresden, Germany

A rat-associated hepevirus, rat hepatitis E virus (ratHEV), was initially detected in Norway rats (*Rattus norvegicus*) from Hamburg, Germany, and subsequently detected in rats from three other cities in Germany and in rats from Vietnam, the US, Indonesia, China, Denmark and France.

Here, we report on a survey of additional Norway rats from Germany, Denmark, France, Switzerland, Austria and Hungary. RatHEV-specific real-time and conventional RT-PCR investigations revealed the presence of ratHEV at the majority of investigated sites in all six countries. In contrast, application of a human pathogenic HEV genotype 1-4-specific real-time RT-PCR did not detect any infections with these genotypes. No difference in age or sex related ratHEV infection status was detected. RatHEV prevalences in towns and rural sites did not differ significantly from high density urban areas, but prevalence in zoological gardens was significantly lower compared to urban areas. Coinfections with rat polyomavirus (ratPyV) and *Leptospira* spp. were detected in 15 of 46 and four of 47 ratHEV-infected rats, respectively. In two rats triple infections with ratHEV, ratPyV and *Leptospira*, all originating from one zoological garden, were detected.

In conclusion, our investigation shows a broad geographical distribution of ratHEV in Norway rats from Europe and its presence in all four habitat types investigated. Future investigations will have to evaluate potential phylogeographical reasons for ratHEV divergence, incursion of ratHEV with invading individuals and to identify putative influences of co-infections by other pathogens to the frequency and outcome of ratHEV infections.

Contact: René Ryll
rene.ryll@fli.de

Virulence determinants of recent avian influenza viruses of subtype H7N7 in chickens

Scheibner D, Veits J, Abdelwhab El-SM, Mettenleiter TC

Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald-Insel Riems, Germany

Avian influenza viruses (AIV), a member of the family Orthomyxoviridae, have a multipartite single-strand RNA genome with eight gene segments coding for at least 10 viral proteins. AIV are classified into 16 hemagglutinin (HA1-HA16) and 9 neuraminidase (NA1-NA9) distinct subtypes. Some H5 and H7 viruses exhibit high virulence in poultry. These highly pathogenic (HP) strains evolve from low pathogenic (LP) progenitors by reassortment and/or point mutations. Changing the monobasic HA cleavage site (CS) in the LP viruses to a polybasic motif is essential to activate HP virus in multiple organs by furin-like proteases causing up to 100 % mortality. In 2015, both LP and HP H7N7 AIVs were isolated from poultry in Germany. Genetic relatedness of both viruses and virulence markers of the HP phenotype in chickens will be investigated using reverse genetics and animal experiments. Therefore, the eight segments of LP and HP viruses will be amplified, cloned in pHWSccdB and used for transfection of HEK293T cells to generate recombinant viruses. In addition to the insertion of a polybasic CS in the LP virus, gene segments of the HP virus will be swapped to determine the minimum constellation that is required for exhibiting high virulence in chickens.

Contact: David Scheibner
david.scheibner@fli.de

Metabolic impact of pneumolysins of *Streptococcus pneumoniae* on human epithelial cells

Schneider S, Lalk M

Institute of Biochemistry, University of Greifswald, Greifswald, Germany

The interplay between the metabolism of human cells and pathogenic bacteria is a crucial pivot-point in infection related processes. Many pathogenic bacteria secrete virulence factors into the medium to gain advances over other organisms and to interfere with the host cell response. To gain deeper insight into physiological adaptations during the contact of *Streptococcus pneumoniae* and eukaryotic cells, an in vitro model will be investigated in which the impact of pneumolysins of *S. pneumoniae* on the metabolism of human epithelial cells will be monitored. Not much is known about the metabolic impact of such virulence factors and a structured metabolome and cell biological study is of particular interest to gain more knowledge about the cellular mechanisms involved in these infections related processes. We will analyze the influence of different pneumolysins on the host cells metabolism. By using bioanalytical techniques like NMR spectroscopy we will investigate the composition of metabolites in extracellular and intracellular samples. In order to connect the metabolic impact to the infection in the host cell methods of multivariate statistical analysis such as PCA and O-PLS will be used. SPME-GC/MS analytics will be employed to analyze lower concentrated compounds and selected volatile metabolites. HPLC methods for the analysis of cellular stress response related metabolites like glutathione are developed in our laboratory and will be used for the investigation of host cell responses. In a future approach we will also include selected primary cell models to analyze the influence of virulence factors like pneumolysis on the epithelial cell metabolism.

Contact: Stefanie Schneider
schneids42@uni-greifswald.de

Development of recombinant live virus vaccines against koi herpesvirus (KHV)

Schröder L,¹ Fuchs W,¹ Bergmann S,² Mettenleiter TC¹

¹Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald-Insel Riems, Germany

²Institute of Infectology, Friedrich-Loeffler-Institut, Greifswald-Insel Riems, Germany

KHV (Cyprinid herpesvirus 3) is a member of the family Alloherpesviridae, which belongs to the order Herpesvirales, and causes a world-wide occurring fatal disease of common carp and koi. To obtain safe live vaccines, we started to generate gene deletion mutants of KHV. Previous studies have shown that the viral thymidine kinase (TK, ORF55) and dUTPase (DUT, ORF123) are dispensable in cell culture, but relevant for virulence in carp. However, attenuation of single mutants based on a KHV field isolate was incomplete, and their in vitro replication was inefficient. Therefore, the TK and DUT genes were now deleted singly and in combination from a cell culture-adapted virus strain (KHV-Taiwan). These mutants grew to high titers, and animal experiments will be done to evaluate their protective efficacy.

To obtain "DIVA" vaccines which permit serological differentiation between immunized and infected fish, we have deleted the genes of two antibody-inducing, but obviously nonessential envelope glycoproteins of KHV (ORF148/149). Another envelope protein of KHV (pORF99) is apparently essential, and trans-complementing cell lines have to be prepared for propagation. The resulting deletion mutant might then be used as a "DISC" vaccine, which is able to replicate in carp, but does not produce infectious virus progeny.

Contact: Lars Schröder
lars.schroeder@fli.de



Interaction of the Foot-and-Mouth Disease Virus with the Innate Immune System

Schult K, Luttermann C

Institute of Immunology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

The Foot-and-Mouth Disease is a highly contagious disease of cloven-hoofed animals. It is characterized by lameness and vesicular lesions on tongue, snout and between cloves. The etiologic agent, the Foot-and-Mouth Disease Virus (FMDV), belongs within the Picornaviridae family to the genus Aphthovirus. Its genome is composed of a single-stranded positive-sense RNA and codes for one polyprotein. The first translated viral protein is the Leaderprotease (LPro) which autocatalytically cleaves itself from the nascent polyprotein. Another protease encoded by the viral genome, the 3C protease (3CPro), cleaves the nascent polyprotein in precursor and mature viral proteins. In addition, both proteases have indicated impact on the transcription and translation machinery and the innate immune response.

To understand the interaction of FMDV with the innate immune system we analyzed the influence of diverse viral proteins on differentially activated IFN-I pathways. Therefore a HEK-293T cell culture system was used and Luciferase reporter assays were performed.

We characterized the LPro mediated inhibition of translation (eIF4G cleavage) and additional specific effects on IFN-I signaling cascades. In our analyses both viral proteases show inhibitory effects on different pathways of IFN-I signaling. Thus, LPro and 3CPro of FMDV interfere with the innate immune system.

To further characterize the interaction of viral proteins with cellular mechanisms more detailed studies are planned to identify the cellular targets of the viral proteins and to clarify the detailed mechanisms of interference with cellular protein translation and the innate immune system.

Contact: [Kristin Schult](mailto:kristin.schult@fli.de)
kristin.schult@fli.de

Newcastle Disease Virus mediates tumor rejection by activation of NK cells but the adaptive immune response prevents relapse in a murine model of pancreatic cancer

Schwaiger T,¹ Knittler MR,¹ Grund C,² Lerch MM,³ Mayerle J,³ Mettenleiter TC,⁴ Blohm U¹

¹Institute of Immunology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

²Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

³Department of Medicine A, University Medicine, Ernst-Moritz-Arndt University Greifswald, Germany

⁴Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Background: Pancreatic cancer is projected to be the 2nd cause of cancer-related deaths worldwide by 2030 and has the worst prognosis of all solid malignancies. In 1957 it was first reported that Newcastle Disease Virus (NDV) has oncolytic properties on tumor cells.

Methods: A single dose of NDV was administered intravenously seven days after orthotopic tumor inoculation of DT6606PDA or Panc02 cells in mice. Tumor growth was measured and immune response was analyzed using flow cytometry. Staining of frozen tumor sections revealed infiltration of leukocytes. Western Blot analysis determined expression of PLC proteins on tumor cells.

Results: A single treatment with NDV inhibited DT6606PDA tumor growth in mice and prevented tumor recurrence for a period of three months. Tumor infiltration and activation of NK cells, cytotoxic and helper T-cells was enhanced. NDV induced melting of Panc02 tumors till d7 pi but recurred henceforth displaying unrestricted tumor growth and active inhibition of tumor-specific immune response. Re-isolated Panc02 cells showed enhanced expression of FoxP3 and TGF- β but neither expression of MHC I nor Rae-1 δ .

Conclusion: NDV is able to reject tumors displaying high immunogenicity and moderate proliferation (DT6606PDA) by mounting an anti-tumor immune response but if non-immunogenic variants accomplish outgrowth (Panc02) anti-tumor response is prevented and relapse occurs. This study underpins the importance of an adaptive immune response which can be initiated by NDV to mediate long-term tumor surveillance apart from direct oncolysis.

Contact: Theresa Schwaiger
theresa.schwaiger@fli.de



Survival of *S. pneumoniae* in sputum of ICU patients.

Seinen J,¹ Dieperink W,² de Smet AMGA,² Hammerschmidt S,¹ van Dijk JM³

¹Department Genetics of Microorganisms, Interfaculty Institute for Genetics and Functional Genomics, Ernst Moritz Arndt University of Greifswald, Germany

²Department of Critical Care, University of Groningen, University Medical Center Groningen, The Netherlands

³Department of Medical Microbiology, University of Groningen, University Medical Center Groningen, The Netherlands

Introduction:

Streptococcus pneumoniae and *Staphylococcus aureus* are known causative agents of ventilator-associated pneumonia (VAP), which leads to increased morbidity and mortality of intensive care unit (ICU) patients. To date, it is not known to which stresses these bacteria are exposed in airways of ventilated patients. The present studies were therefore aimed at establishing an ex vivo assay that mimics the in vivo situation through incubation of bacteria in sputum samples collected from different ICU patients.

Material and methods:

Wild-type *S. pneumoniae* TIGR4 and different virulence-attenuated mutants were incubated in sputum samples from mechanically ventilated ICU patients from the University Medical Center Groningen. Next, bacterial survival in the sputum samples was tested by plating. *S. pneumoniae* was also used in a sputum spotting assay, where sputum was spotted on a blood agar plate and the inhibition zones were compared to those observed for *Streptococcus anginosus* and *S. aureus*.

Results and Conclusions:

The tested *S. pneumoniae* strains showed different rates of survival in sputa from different patients. Intriguingly, the spotting assay revealed major differences in the bactericidal activity of sputa from different patients. Our ongoing research is aimed at assessing both patient parameters that affect bacterial survival in sputum as well as the bacterial gene repertoire needed for survival.

Contact: Jolien Seinen
j.seinen@umcg.nl



Macrophage cell death during infection with *Burkholderia pseudomallei*

Stiehler J,¹ Bast A,¹ Steinmetz I^{1,2}

¹University Medicine Greifswald, Friedrich Loeffler Institute of Medical Microbiology, Greifswald, Germany

²Medical University of Graz, Institute of Hygiene, Microbiology and Environmental Medicine, Graz, Austria

Macrophage cell death is an intrinsic immune defense mechanism in response to microbial infection. It can remove the replicative niche of intracellular pathogens and induces immune attack. However, uncontrolled cell death likely contributes to tissue damage, which may enhance bacterial dissemination. Apoptosis, necroptosis and pyroptosis constitute the three major cell death modes and are tightly regulated to ensure adequate immune reactions to invading pathogens.

We recently provided evidence that infection of primary murine macrophages with the cytosolic pathogen *Burkholderia pseudomallei* and causative agent of melioidosis leads to early induction of caspase-1/11-dependent pyroptosis, a highly inflammatory and lytic cell death. We identified the *B. pseudomallei* type three secretion system (T3SS) rod protein as a stimulator of caspase-1. In the absence of caspase-1 and -11 delayed activation of alternate cell death pathways such as inflammatory silent apoptosis might play a role during infection. This project will focus on additional caspase-1/11-independent cell death pathways in macrophages. We want to clarify the contribution of apoptotic caspases to resistance against *B. pseudomallei*. This study aims to identify and characterize bacterial factors leading to cell death induction. Finally, we will analyze the crosstalk between different forms of macrophage death.

Contact: [Julia Stiehler](mailto:julia.stiehler@uni-greifswald.de)
julia.stiehler@uni-greifswald.de

Proteome profiling of *Burkholderia pseudomallei* quorum sensing mutants

Stoll-Ziegler K,¹ Kohler C,² Zühlke D,¹ Ziesche L,³ Steinmetz I,² Riedel K¹

¹University of Greifswald, Institute of Microbiology, Greifswald, Germany

²University Medicine Greifswald, Friedrich Loeffler Institute of Medical Microbiology, Greifswald, Germany

³TU Braunschweig, Institute of Organic Chemistry, Braunschweig, Germany

Burkholderia pseudomallei, the causative agent of melioidosis, is a Gram-negative soil bacterium found in tropical areas. *B. pseudomallei* employs several acyl-homoserine lactone (AHL)-mediated quorum sensing (QS) systems, which activate specific sets of genes depending on cell density [1,2]. The genome of *B. pseudomallei* encodes genes for three QS systems with one luxI and one luxR homologue, respectively, and additionally three orphan luxR homologues [3]. The luxI homologues encode AHL-synthases producing specific AHLs binding to their respective luxR-coded transcriptional regulator, which in turn regulates the expression of virulence-related genes [4]. The aim of this study was to dissect the impact of the different QS systems on gene expression and pathogenicity of *B. pseudomallei*.

As a starting point single, double and triple deletion mutants were constructed and sub-proteome fractions of wild-type and mutant strains were subjected to mass spectrometry analysis aiming on the identification of QS-target genes. Cytosolic proteins were identified by the DIA approach IMSE, whilst extracellular and surface-associated proteins were analyzed by GeLC MS/MS. Moreover, the AHL-spectrum of the different mutants was analyzed by mass spectrometry and the mutants were characterized with respect to different phenotypes. Our comprehensive proteome analysis revealed a significant number of QS-regulated genes, some of which have been already described as QS-regulated in other Gram-negative opportunistic pathogens, e.g. *Burkholderia cenocepacia* [5] and are known to be involved in pathogenicity. We are currently investigating the virulence potential of the QS-mutants in different pathogenicity models.

Contact: Karolin Stoll-Ziegler
stoll-ziek@uni-greifswald.de



Optimizing murine infection models by using mouse-adapted *S. aureus* strains

Trübe P,¹ Hertlein T,² Krause B,¹ Zeun J,¹ Mrochen D,¹ Bröker B,¹ Ohlsen K,² Holtfreter S¹

¹Department of Immunology, University Medicine Greifswald, Germany

²Institute for Molecular Infection Biology, University of Würzburg, Germany

In general, to study the pathogenicity of *S. aureus* human-adapted *S. aureus* strains are used in murine infection models. However, *S. aureus* is highly host-specific. Several virulence factors, such as PVL, CHIPS, SCIN, are human-specific and require non-physiological concentrations in the mouse model or do not work at all. We have recently reported that mice are natural hosts of *S. aureus*. Our data show that murine *S. aureus* strains belong to unique clonal clusters and seem to be well adapted to their host. The use of such mouse-adapted *S. aureus* strains may be a promising tool to develop more clinically relevant infection models.

To determine whether mouse-adapted *S. aureus* strains can be used to optimize murine infection models, *S. aureus* strains isolated from wild and laboratory mice were compared to the human *S. aureus* strain Newman. We used two different infection models, pneumonia and bacteremia, and analyzed the survival and the bacterial load in several organs.

There were significant differences in the survival rate between the strains in both infection models. Furthermore, the bacterial load within the organs differed between the strains. These results indicate that mouse-adapted *S. aureus* strains have a better fitness in murine infection models and can cause more severe disease than *S. aureus* Newman. Notably, a CC49 isolate from a wild mouse was highly virulent in laboratory mice and is in the focus of ongoing studies. With our study we hope to improve mouse infection models that are required to study *S. aureus* - host interaction.

Contact: Patricia Trübe
patricia.truebe@uni-greifswald.de

Hyperfusogenic mutations in the herpesvirus fusion protein – in search of a super-fuser

Vallbracht M,¹ Schröter C,¹ Klupp BG,¹ Kühn J,² Mettenleiter TC¹

¹Institute of Molecular Virology and Cell Biology, Friedrich-Loeffler-Institut, Greifswald – Insel Riems, Germany

²University of Münster, Institute of Medical Microbiology – Clinical Virology, Münster, Germany

Enveloped viruses utilize membrane fusion to infect and spread in their hosts. Entry is initiated by binding of the virus to specific cellular receptors and fusion with the cellular membrane is catalyzed by a viral fusion protein. While many viruses encode only one protein capable of performing attachment and fusion, herpesviruses require four different proteins: a receptor-binding protein and the conserved fusion-machinery composed of glycoprotein (g)B and the gH/gL complex. Although gB possesses typical features of a class III fusion protein, it is unable to execute membrane fusion on its own but depends on gH/gL. However, although gB and gH/gL are crucial for entry, the alphaherpesvirus Pseudorabies Virus (PrV) is able to perform limited cell-to-cell spread in absence of gL. Passaging experiments of gL-deleted PrV resulted in a mutant which efficiently entered cells in absence of gL. Among others, this mutant carries mutations in gB (gBB4.1) resulting in efficient fusion with only gH.

Aim of the present study was to investigate whether additional amino acid changes might convert gB into an autonomic fusion protein. For this purpose, we introduced additional mutations into PrV gBB4.1 which had been described to enhance fusogenicity of herpes-simplex and varicella-zoster viruses. To test the activity of these mutants we used a transfection-based fusion assay. Our results show that introduction of additional mutations into gBB4.1 further enhanced fusogenicity. Despite the excessive fusion-activity, this gB mutant still depends on gH, indicating that raising the fusogenic potential is not sufficient to compensate for gH function.

Contact: Melina Vallbracht
Melina.Vallbracht@fli.bund.de



Cell-surface localized pneumococcal proteins – new insights into the abundance and immunogenicity

Voß F,¹ Sundaramoorthy N,² Abdullah M,¹ Michalik S,² Saleh M,¹ Schmidt F,² Hammerschmidt S¹

¹Department Genetics of Microorganisms, Interfaculty Institute for Genetics and Functional Genomics, Ernst Moritz Arndt University of Greifswald, Germany

²ZIK-FunGene Junior Research Group "Applied Proteomics", Department of Functional Genomics, Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald, Greifswald, Germany

Question and aims. Pneumococcal next-generation vaccines would be serotype-independent and protein-based formulations. The pneumococcal cell surface is decorated with several clusters of proteins, namely choline-binding proteins (CBP), sortase anchored or (trans-)membrane associated proteins, lipoproteins and non-classical surface proteins. Highly conserved proteins from different clusters representing potential candidates for an innovative vaccine were analyzed regarding their abundance on the pneumococcal surface and immunogenicity.

Methods. The cell-surface abundance of proteins was examined by flow cytometry using protein specific polyclonal IgGs generated in mice. The relative antibody titers from these mouse sera were determined by ELISA and antibody titers were also analyzed with the Luminex® FlexMap3D® technique. In addition, the immunogenicity of the selected proteins was screened with convalescent sera from patients infected with different pneumococcal serotypes.

Results. The Luminex-based immunoproteomics approach showed that nearly all proteins are immunogenic when administered to mice. Analysis of the convalescent patient sera revealed 5 proteins from different surface protein clusters which induced high antibody titers during pneumococcal infections. These proteins most likely also showed a high abundance on the pneumococcal cell surface in flow cytometry. Interestingly nearly all antibodies also bound to the encapsulated strain D39.

Conclusion. The analyzed pneumococcal surface proteins are highly conserved and immunogenic. Besides, surface abundance of some proteins is high and partly correlates with the observed immunogenicity. Therefore, these proteins represent promising candidates for a protein-based conjugate or subunit vaccine, which are urgently needed to combat pneumococcal infections.

Contact: Franziska Voß
franziska.voss@uni-greifswald.de



Identification and molecular characterization of highly immunogenic mycobacterial lipopeptides

Woelke S, Bastian M

Independent Research Group for Vaccines and Vaccination Safety, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

A number of recent publications describe mycobacterial lipopeptides as a novel class of highly immunogenic lipid antigens in mycobacteria infected humans (Seshadri et al., JI, 2013; Bastian et al., JI, 2008), bovines (Van Rhijn et al., EJI, 2009) and guinea pigs (Kaufmann et al., JI, 2016). Previous data of our group show, that these lipopeptides are presented by MHCII and induces a strong CD4+ T-lymphocytes immune reaction in TB-infected or BCG vaccinated individuals. We could demonstrate that the reaction of these lymphocytes is specific for mycobacteria of the Mycobacterium Tuberculosis Complex (MTC).

The aim of the presented PhD project is to identify the molecular identity of these antigens. To this end we will establish mono- or oligoclonal T-lymphocyte lines of PPD reactive humans and/or BCG sensitized guineapigs.

In parallel, we will fractionate whole lipid extracts of M.tuberculosis using phase separation and column chromatography techniques.

With the antigen specific T-cell lines we will test the obtained fractions and will identify those that contain highly immunogenic lipopeptides.

In a next step we will characterize the molecular identity of the antigens.

The identification of the molecular nature of the immunogenic lipopeptides is a prerequisite to further elucidate the role of the antigens in the fine-tuned host pathogen interplay. In addition, it will be assessed to what extent these antigens contribute to the Tuberculin-Skin-Reaction, i.e. whether they can be exploited as diagnostic antigens , or whether they could serve as components of a future TB-vaccine.

Contact: Sören Woelke
Soeren.Woelke@fli.de

Establishment of an antibiotic signature library for *Staphylococcus aureus*

Wolff C, Zühlke D, Pané-Farré J, Riedel K

Institute of Microbiology, Ernst-Moritz-Arndt-University Greifswald, Greifswald, Germany

Worldwide spread of antibiotic resistance greatly impairs treatment of bacterial infections. Therefore, antibacterial agents with new mechanisms of action and new innovative and cost-effective techniques for the characterization and validation of new drugs are urgently needed. Gel-based and gel-free proteomics have emerged as valuable tools to study the physiology of microbes under antibiotic stress conditions. Our study aims at the creation of a comprehensive antibiotic signature library for *Staphylococcus aureus*. Protein signatures of *S. aureus* treated with well characterized antibiotics will be used to identify cellular targets and the mode-of-action of new anti-microbial compounds.

S. aureus HG001 was exposed to various concentrations of antibiotics with well-defined molecular targets to determine their minimal inhibitory concentrations (MIC). Multiples of the MICs were tested in growth experiments to identify antibiotic concentrations that reduced the bacterial growth but did not inhibit growth completely in order to map the specific antibiotic stress response. Cytosolic protein extracts of *S. aureus* HG001 treated with antibiotics were analyzed using a gel-free LC-IMSE approach in combination with the Hi3 method for absolute protein quantification.

Although each antibiotic showed an individual protein expression profile, signature proteins specific for a common drug target were identified (e.g. cell wall, ribosome, replication machinery). These subsets of proteins whose expression levels are characteristic for a specific antibiotic treatment have been designated as "proteomic signature".

We established a workflow for a comprehensive antibiotic stress proteome signature library of *S. aureus*. Using this pipeline, new compounds can be evaluated to gain insight into their mode-of-action.

Contact: Christian Wolff
christian.wolff@uni-greifswald.de



Immune mechanisms involved in anti-viral immune response of European bats against relevant Lyssaviruses

Zhu Y, Schäfer A, Köllner B

Institute of Immunology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Lyssaviruses are the causative agents of rabies, a fatal zoonotic disease of nerve system. European bats harbor two specific types of Lyssaviruses, European Bat Lyssavirus (EBLV-1 and -2). Although cases of rabies caused by EBLV 1 or 2 are described there are no reports about epidemics in bats. Due to the long co-evolution between bats and EBLVs innate immune pathways might contribute to this disease resistance in European bats.

We focus on specific physiology of bats, which might lead to their 'special' immune response against lyssaviruses. Using established cell lines from *M. myotis* (*M. myotis* brain (MmBr), nervus olfactorius (MmNol), and nasal epithelium (MmNep) we simulate the transmission of lyssaviruses along the aerosol infection route. The anti-viral effects, induction of interferons (IFNs)/interferon stimulated genes (ISGs) and expressions of viral receptors in each cell line will be investigated.

Bats are the only mammals which actively fly. Their flight ability enables a higher body temperature. High body temperatures ranges increase the rate of multiple immune responses in bats, and might stimulates innate immune response, such as IFNs to repress lyssavirus replication. Using *M. myotis* cell lines we investigate lyssavirus replication in different temperatures to simulate bat's daily activity and their hibernation in vitro. We build growth curve to find different sensitivities of cell lines, simulate RNA-viral infection with Toll like receptor (TLR) agonists, and incubate cell lines in aerosol infection pathway by 39°C, 37°C, 25°C, to simulate the daily activity of bats, and by 4°C to simulate the hibernation of bats.

Contact: [Yaqing Zhu](mailto:yaqing.zhu@fli.de)
yaqing.zhu@fli.de

Identification of gene signatures associated with foot-and-mouth disease virus infection by transcriptome analysis

Zoli M, Höper D, Dill V, Beer M

Institute of Diagnostic Virology, Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany

Foot-and-mouth disease virus (FMDV) is the etiological agent of a dramatic disease that affects cloven-hoofed animals. The pathogenesis of this disease is nowadays still not fully understood; in particular, it is unknown why some animals become persistently infected, despite being vaccinated. Therefore, there is the need to identify at the transcriptome level both virus and host genes involved in FMDV acute and persistent infection, in order to find the target genes to develop novel vaccines. Transcriptome shotgun sequencing (RNA sequencing) has rapidly become the leading tool to study how the transcriptome evolves, in particular how the genes are differentially expressed during virus infection. In a time series experiment, BHK-21 cells have been infected with FMDV serotype O1 Manisa at a multiplicity of infection (MOI) of 1, microscopically monitored for cytopathic effect and samples have been collected at different times post infection. From each time point, the total RNA was extracted and the poly(A)mRNA was isolated; the latter was subsequently used to establish cDNA libraries. For each library, RNA sequencing was performed both with Ion Torrent PGM and with Illumina MiSeq platforms. The differentially expressed genes have been identified statistically among the chosen time points. Through the differentially expressed genes we have found genes coding for proteins involved in viral replication, apoptotic processes, protein localization and transport, regulation of cell growth and cellular responses to various stress stimuli. In order to confirm the transcriptome results, differentially expressed genes will be selected for further investigations.

Contact: [Martina Zoli](mailto:martina.zoli@fli.de)
martina.zoli@fli.de

Impressum

Friedrich-Loeffler-Institut, Greifswald - Insel Riems, Germany
Ernst-Moritz-Arndt-Universität Greifswald, Germany
Alfried Krupp Wissenschaftskolleg Greifswald, Germany

Edited by: Thomas C. Mettenleiter, Friedrich-Loeffler-Institut
Sven Hammerschmidt, Ernst-Moritz-Arndt-Universität
Bärbel Friedrich, Alfred Krupp Wissenschaftskolleg Greifswald
Sylvia Kohler, Ernst-Moritz-Arndt-Universität
Bianca M. Bussmann, Friedrich-Loeffler-Institut
Composition & Layout: Katja von Einsiedel, Friedrich-Loeffler-Institut
Bianca M. Bussmann, Friedrich-Loeffler-Institut
Print: Bundesamt für Seeschifffahrt und Hydrographie

Greifswald, September 2016

