

F1 and Tbilisi Are Closely Related Brucellaphages Exhibiting Some Distinct Nucleotide Variations Which Determine the Host Specificity

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We report on the 41,143-bp genome of brucellaphage F1, a podovirus that infects several *Brucella* species. The F1 genome is almost identical to the genome of brucellaphage Tb. However, some structural proteins of the phages exhibit extensive polymorphisms and might be responsible for their different host ranges.

Received 23 December 2013 Accepted 31 December 2013 Published 30 January 2014

Citation Hammerl JA, Al Dahouk S, Nöckler K, Göllner C, Appel B, Hertwig S. 2014. F1 and Tbilisi are closely related brucellaphages exhibiting some distinct nucleotide variations which determine the host specificity. Genome Announc. 2(1):e01250-13. doi:10.1128/genomeA.01250-13.

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Brucellae are facultative intracellular pathogens that may cause reproductive failure and abortion in animals and a feverish multiorgan disease in humans (1). Brucellaphages have been used as a diagnostic tool since the 1960s, when phage Tbilisi (Tb) was discovered (2). According to their host specificities, seven *Brucella* phage groups (prototypes Tb, Fi, Wb, Bk2, R/C, Iz, and Np) (3, 4) have been established, even though various host ranges have been suggested for phage Tb (3–5). The currently known brucellaphages share a podoviral morphology (6) and strong DNA homologies (4). However, only two brucellaphages (Tb and Pr) have been sequenced so far (7).

Here, we report the complete nucleotide sequence of phage F1 (8), which is routinely used for Brucella typing. F1 phage stocks were provided by the World Organisation for Animal Health (OIE) Brucellosis Reference Center of the Veterinary Laboratories Agency (Addlestone, United Kingdom). The phages were propagated on the Brucella abortus vaccine strain S19 and purified according to standard procedures (9). Similar to the host range of Tb reported by Corbel (3), F1 was propagated on B. abortus, Brucella suis (bv1 and bv5), Brucella neotomae, and Brucella microti reference strains. The plaques formed by F1 were large (2 mm in diameter) and clear. Like other brucellaphages, F1 has the typical morphology of a podovirus. Phage DNA was isolated by proteinase K/SDS treatment of CsCl-purified particles as previously described (9). Whole-genome sequencing was performed with the 454 genome sequencer FLX Titanium system (Roche, Germany). Assembling of reads (Newbler Assembler version 2.6) yielded a single linear contig with a >30-fold sequence coverage. Gene prediction and annotation of the genome sequence were carried out using MyRAST (10-12).

Phage F1 has a linear double-stranded genome of 41,143 bp, with an average G+C content of 48.2%. Bal31 analyses revealed that the genome is circularly permuted. Fifty-eight putative genes and seven transcriptional terminators have been identified. A functional assignment was made for 17 gene products. F1 is very similar to the sequenced brucellaphages Tb and Pr, with nucleo-tide identities of 99% and 98%, respectively. Most of the predicted gene products are even 100% identical. However, some F1 prod-

ucts that are presumably involved in virion assembly (gp12, major head protein; gp15 and gp16, structural proteins; gp24, tail spike protein) show striking amino acid polymorphisms in relation to the corresponding Tb proteins. Notably, Flores et al. (7) reported that the sequenced Tb phage exclusively infects *B. abortus* strains. Thus, it is very likely that the diverging structural proteins of F1 and Tb account for the diverging host range of the phages. Nevertheless, the high rates of identity suggest that the hithertodescribed brucellaphages are host range mutants originating from a common ancestor. Since only a small number of F1 products diverge from Tb and Pr, it should soon be possible to identify the amino acids that are important for host specificity. This work can be facilitated by sequencing of brucellaphages that belong to other host range groups.

Nucleotide sequence accession number. The genome of brucellaphage F1 is available under the Genbank accession no. HG428758.

ACKNOWLEDGMENTS

The work of J.A. Hammerl was supported by a grant of the Federal Institute for Risk Assessment (BfR 1332-488) and the German Federal Ministry of Education and Research (SiLeBAT, project no. 13N11202).

REFERENCES

- 1. Godfroid J, Al Dahouk S, Pappas G, Roth F, Matope G, Muma J, Marcotty T, Pfeiffer D, Skjerve E. 2013. A "One Health" surveillance and control of brucellosis in developing countries: moving away from improvisation. Comp. Immunol. Microbiol. Infect. Dis. 36:241–248. http://dx .doi.org/10.1016/j.cimid.2012.09.001.
- 2. Drozevkina MS. 1963. The present position in *Brucella* phage research. Bull. World Health Organ. **29**:43–57.
- Corbel MJ. 1987. Brucella phages: advances in the development of a reliable phage typing system for smooth and non-smooth Brucella isolates. Ann. Inst. Pasteur Microbiol. 138:70–75. http://dx.doi.org/10.1016/0769 -2609(87)90056-1.
- 4. Rigby CE, Cerqueira-Campos ML, Kelly HA, Surujballi OP. 1989. Properties and partial genetic characterization of Nepean phage and other lytic phages of *Brucella* species. Can. J. Vet. Res. 53:319–325.
- Taran IF, Zanina VM, Liamkin GI, Tsybin BP, Tikhenko NI. 1983. Comparative evaluation of the spectrum of lytic effects of bacteriophages Tb, Wb, Fi, Bk2 and R on various *Brucella* species. Zh. Mikrobiol. Epidemiol. Immunobiol. 2:48–52. (Article in Russian.)

- Ackermann HW, Simon F, Verger JM. 1981. A survey of *Brucella* phages and morphology of new isolates. Intervirology 16:1–7. http://dx.doi.org/ 10.1159/000149240.
- Flores V, López-Merino A, Mendoza-Hernandez G, Guarneros G. 2012. Comparative genomic analysis of two brucellaphages of distant origins. Genomics 99:233–240. http://dx.doi.org/10.1016/j.ygeno.2012.01.001.
- Calderone JG, Pickett MJ. 1965. Characterization of brucellaphages. J. Gen. Microbiol. 39:1–10. http://dx.doi.org/10.1099/00221287-39-1-1.
- 9. Sambrook J, Russel D. 2001. Molecular cloning: a laboratory manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T,

Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/1471-2164-9-75.

- 11. Glass EM, Wilkening J, Wilke A, Antonopoulos D, Meyer F. 2010. Using the metagenomics RAST server (MG-RAST) for analyzing shotgun metagenomes. Cold Spring Harb. Protoc. 2010:pdb.prot5368. http://dx.doi .org/10.1101/pdb.prot5368.
- Meyer F, Paarmann D, D'Souza M, Olson R, Glass EM, Kubal M, Paczian T, Rodriguez A, Stevens R, Wilke A, Wilkening J, Edwards RA. 2008. The metagenomics RAST server—a public resource for the automatic phylogenetic and functional analysis of metagenomes. BMC Bioinformatics 9:386. http://dx.doi.org/10.1186/1471-2105-9-386.